

**Can multivariate curve  
resolution be used for  
quantitative purposes?**

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# Outline

- **Introduction to MCR-ALS method**
- Quantitative MCR-ALS for two-way data
- Quantitative MCR-ALS for three-way data
- Conclusions and Acknowledgements

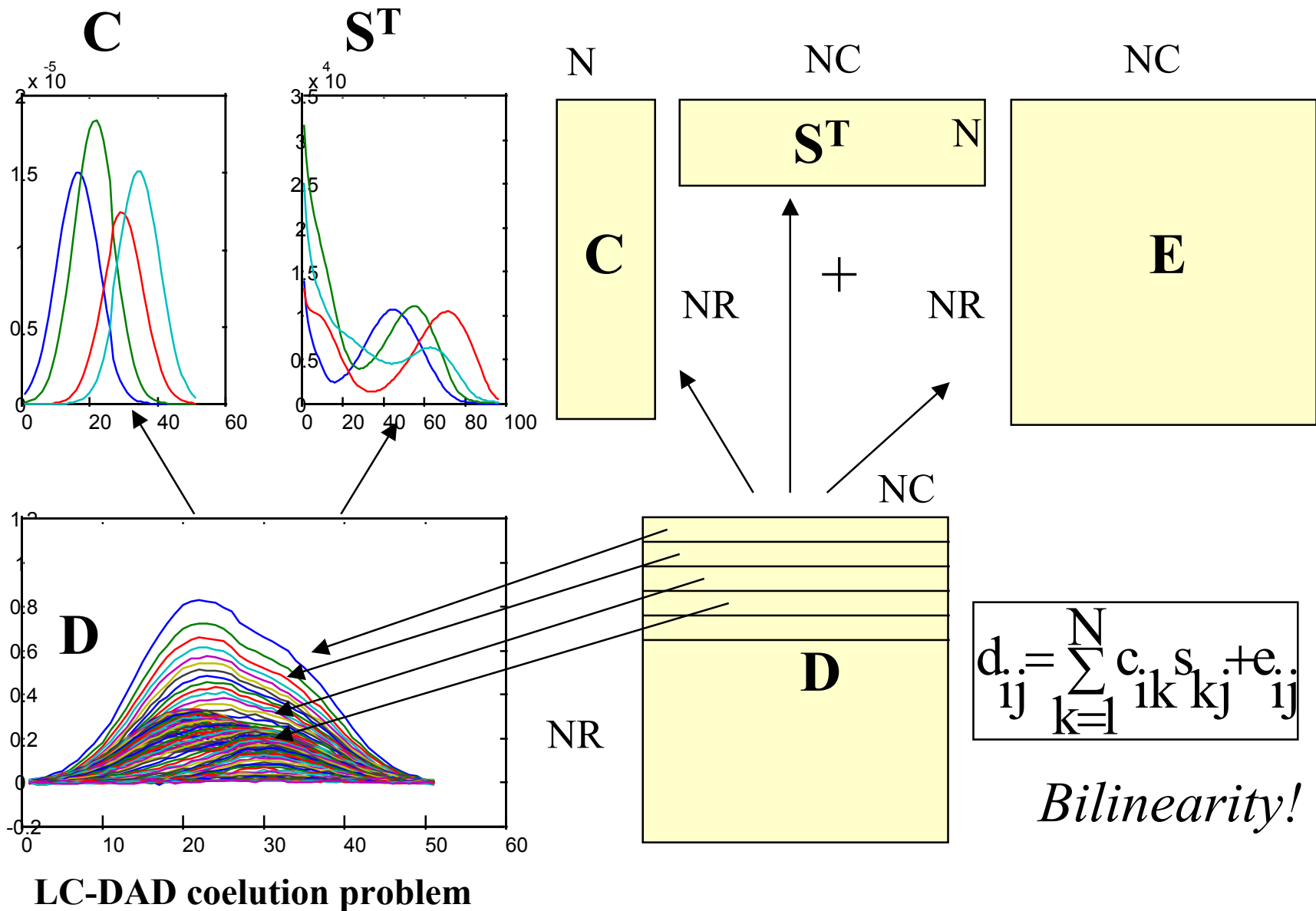
# Multivariate Self Modeling Curve Resolution

Group of techniques which intend the **recovery of the response profiles** (spectra, pH profiles, time profiles, elution profiles,...) of more than one component in an **unresolved and unknown mixture** (*obtained from evolutionary processes*) when **no prior or little information is available** about the nature and composition of these mixtures.

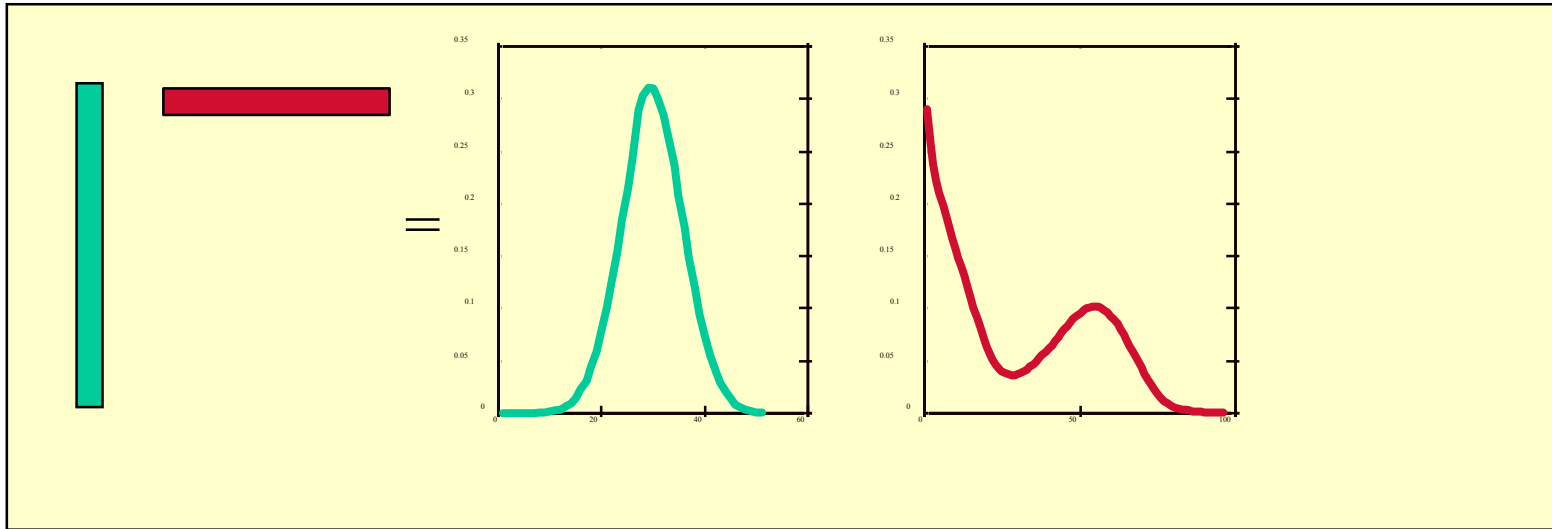
# What problems can be solved with MCR?

- **Mixture Analysis problems using multivariate detection methods (spectrometry, voltamperometry,...)**
- **Chromatographic coelution problems**
- **Industrial process monitoring and process mixture analysis problems**
- **Resolution of chemical reaction based systems (equilibria and kinetics)**
- **Resolution of macromolecular conformations; polynucleotide chemistry; protein folding**
- **Resolution of environmental pollution sources**
- **.....**

# Multivariate Curve Resolution



# GOALS OF MCR



- **Recovery of the responses of every component (chemical species) in the different orders of measurement**
- **Is it possible to recover quantitative information?**

# Rotational Ambiguities

$$\mathbf{D} = \mathbf{C} \mathbf{S}^T + \mathbf{E} = \mathbf{D}^* + \mathbf{E}$$

$$\mathbf{S}_{\text{new}}^T = \mathbf{T} \mathbf{S}^T$$

$(N, NC)$     $(N, N)$     $(N, NC)$

$$\mathbf{C}_{\text{new}} = \mathbf{C} \mathbf{T}^{-1}$$

$(NR, N)$     $(NR, N)$     $(N, N)$

$$\mathbf{D}^* = \mathbf{C} \mathbf{S}^T = \mathbf{C} \mathbf{T}^{-1} \mathbf{T} \mathbf{S}^T = \mathbf{C}_{\text{new}} \mathbf{S}_{\text{new}}^T$$

**Matrix decomposition is not unique!**

$\mathbf{T}(N, N)$  is any non-singular matrix

Rotational freedom for any  $\mathbf{T}$

# How to break (*at least partially!*) rotational ambiguities?

- ✓ By using natural and shape constraints
- ✓ By using selective variables and/or local rank information (equality constraints)
- ✓ By matrix augmentation (three-way data analysis)

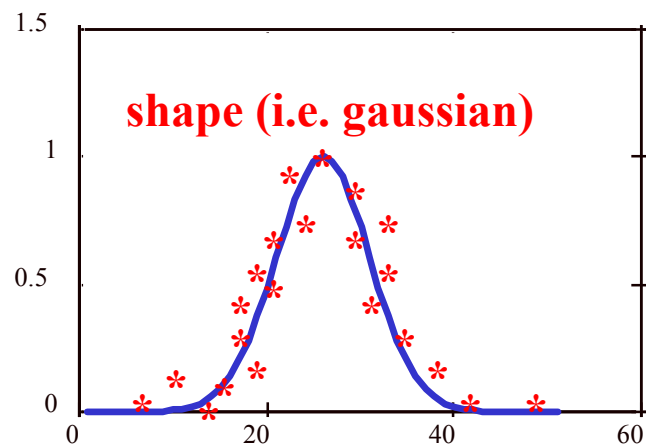
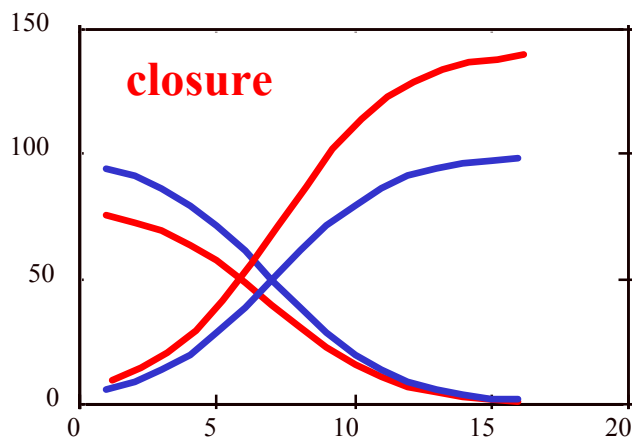
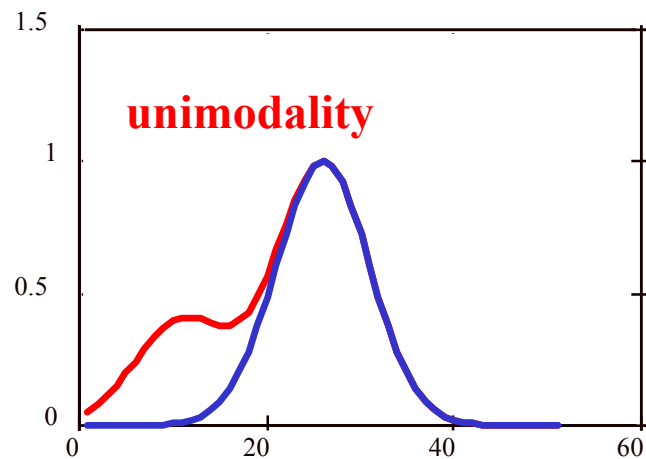
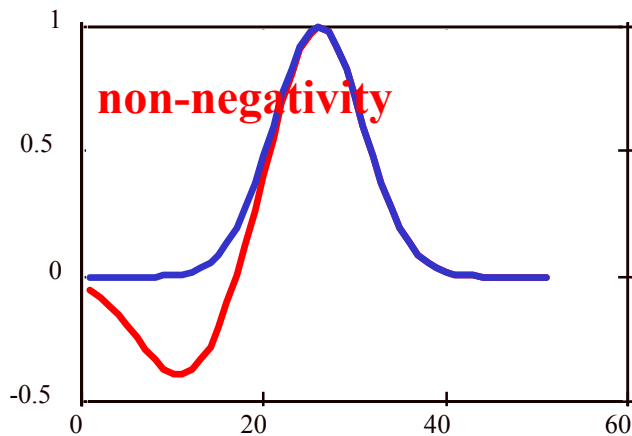
*Tauler R, Izquierdo-Ridorsa A. and Casassas E., Chemom. Intell. Lab. Systems, 1993, 18, 293-300.*

*Tauler R. , Smilde A. and Kowalski B. R., J.of Chemometrics, 1995,9,31*

*Tauler R. Chemom.Intell.Lab.Sys., 1995, 30, 133*



# Natural and shape constraints (graphical explanation)

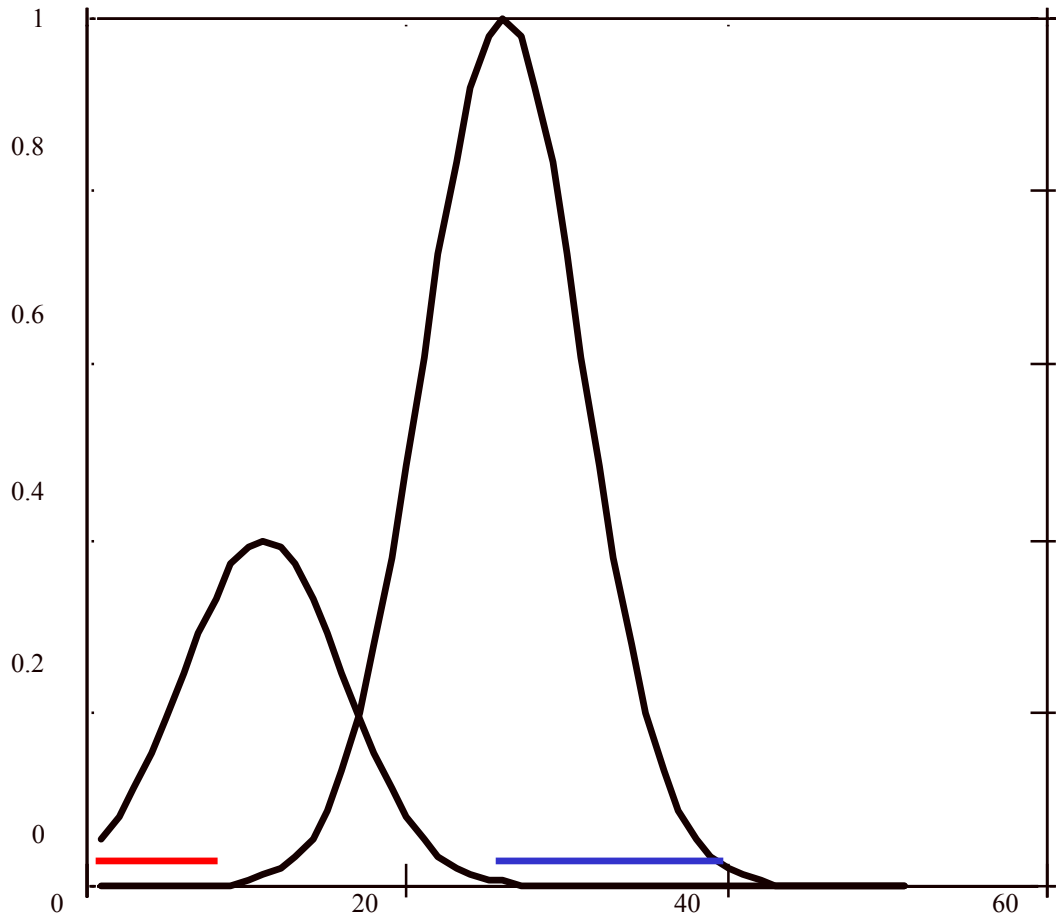


# Selectivity constraints

**Selective regions are  
regions where only  
one component is present**

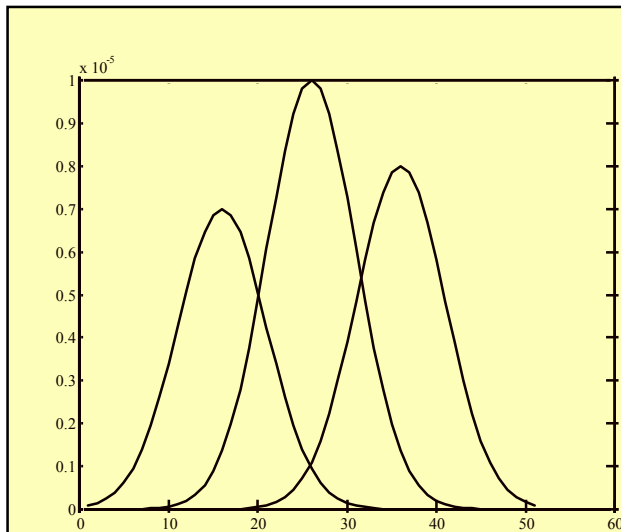


**In these regions  
rotational  
ambiguities are solved  
easily**

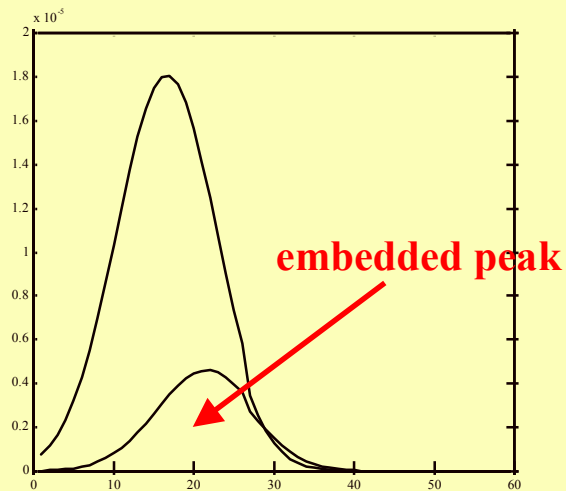


# Resolution Based on Local Rank Information

*Rolf Manne, Chemometrics and Intelligent Laboratory Systems, 27, 1995, 89-94*

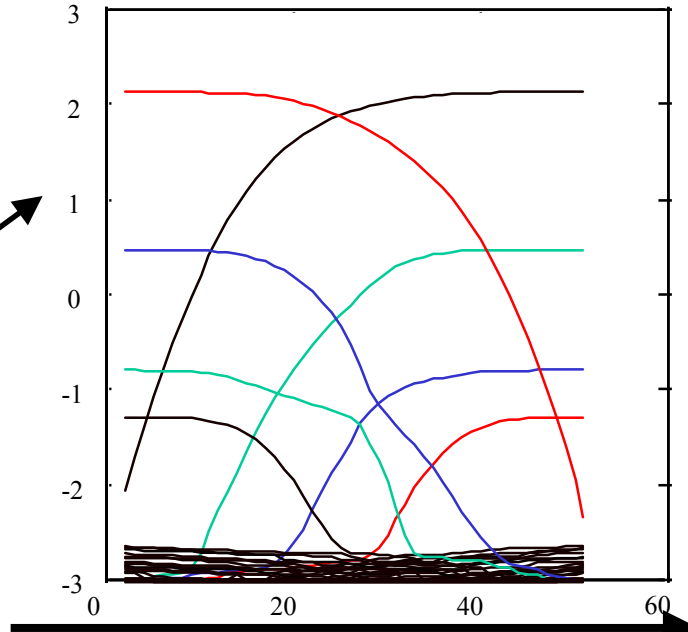
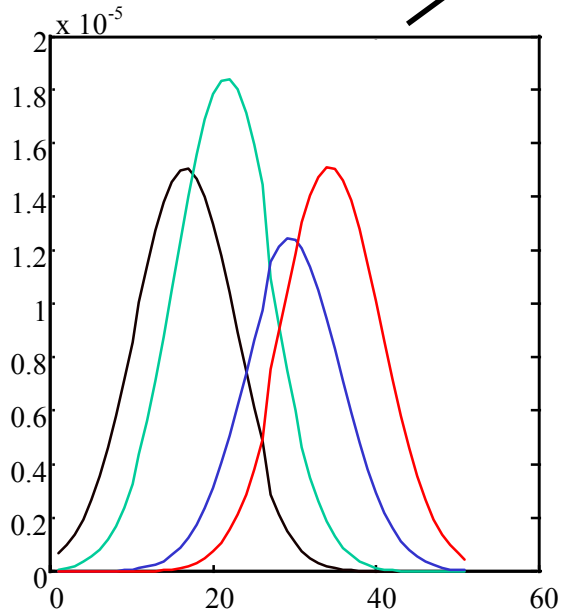


**this system may be resolved  
without ambiguities**



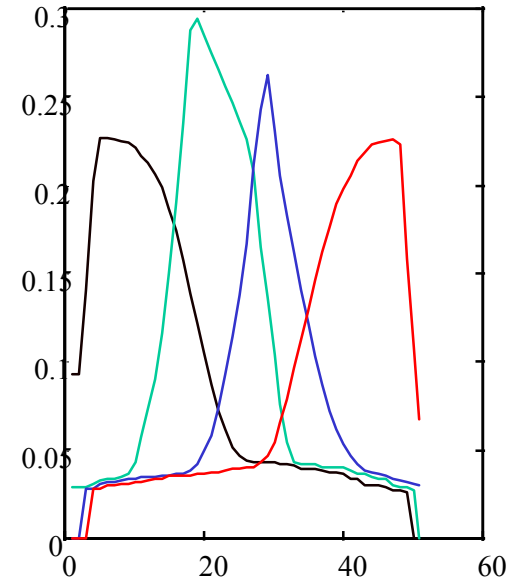
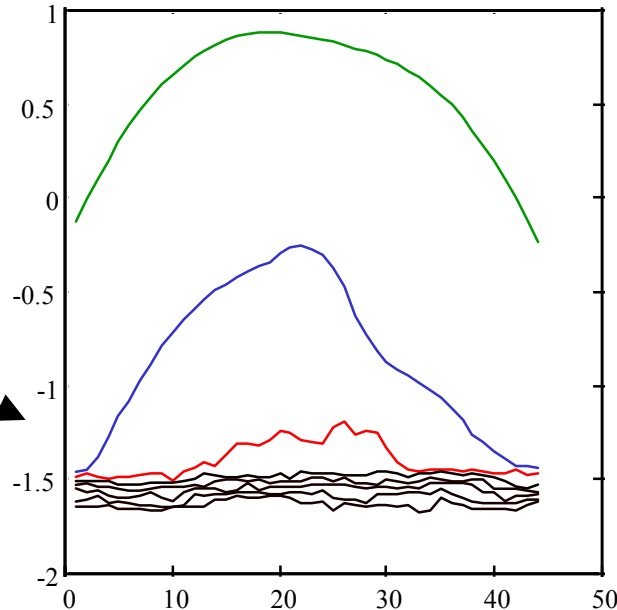
**this system cannot be resolved  
without ambiguities**

# Selectivity and Local Rank detection **EFA**

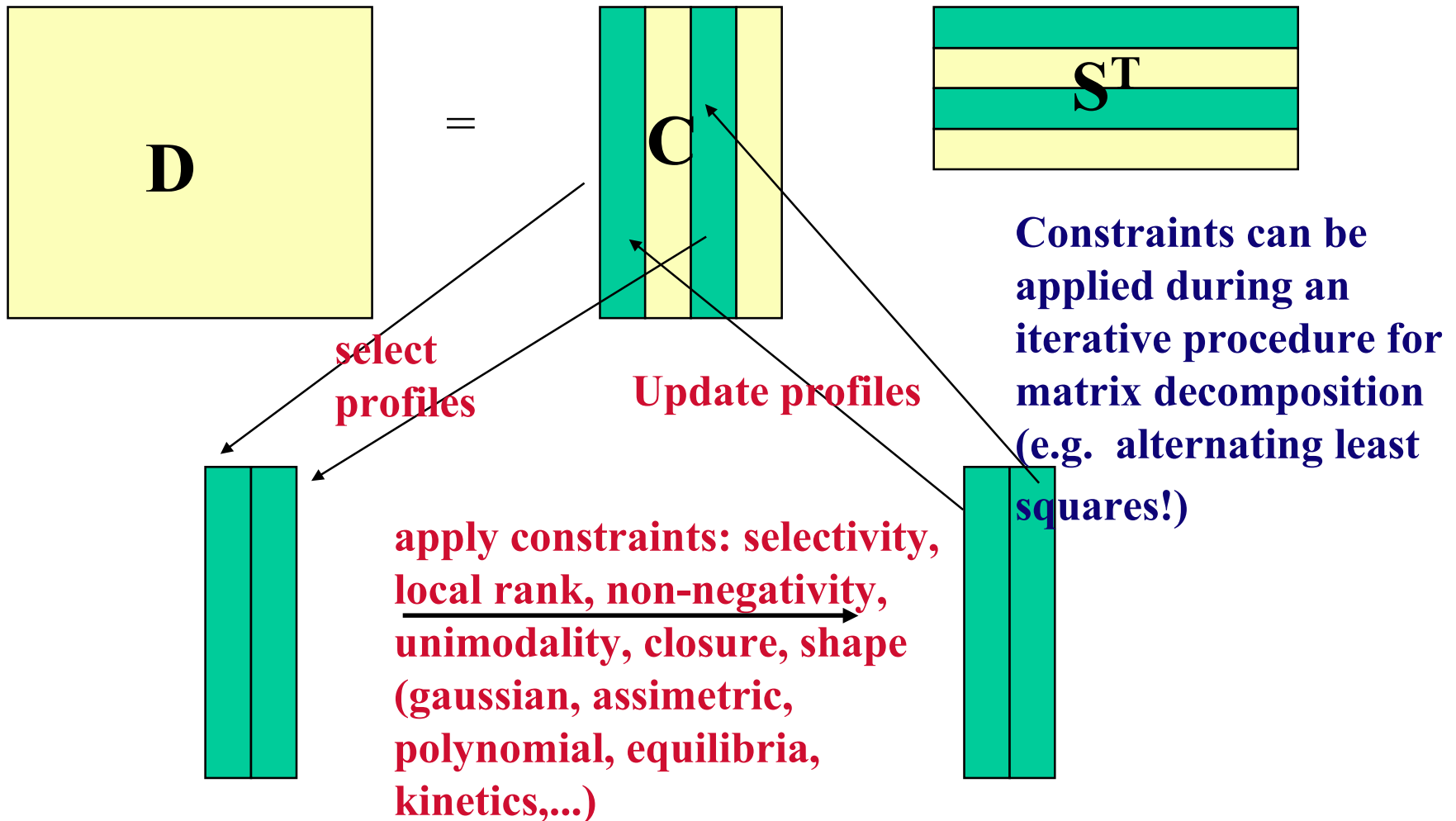


**initial estimates**

**FSMWEFA**



# Application of constraints



# Multivariate Curve Resolution Alternating Least Squares (MCR-ALS)

## *Alternating Least Squares (ALS) solution*

- Initial estimates of  $C$  or  $S^T$  are obtained from EFA or from pure variable detection methods
- **Optional constraints are applied at each ALS iteration !**

$$\mathbf{D} = \mathbf{C} \mathbf{S}^T + \mathbf{E}$$

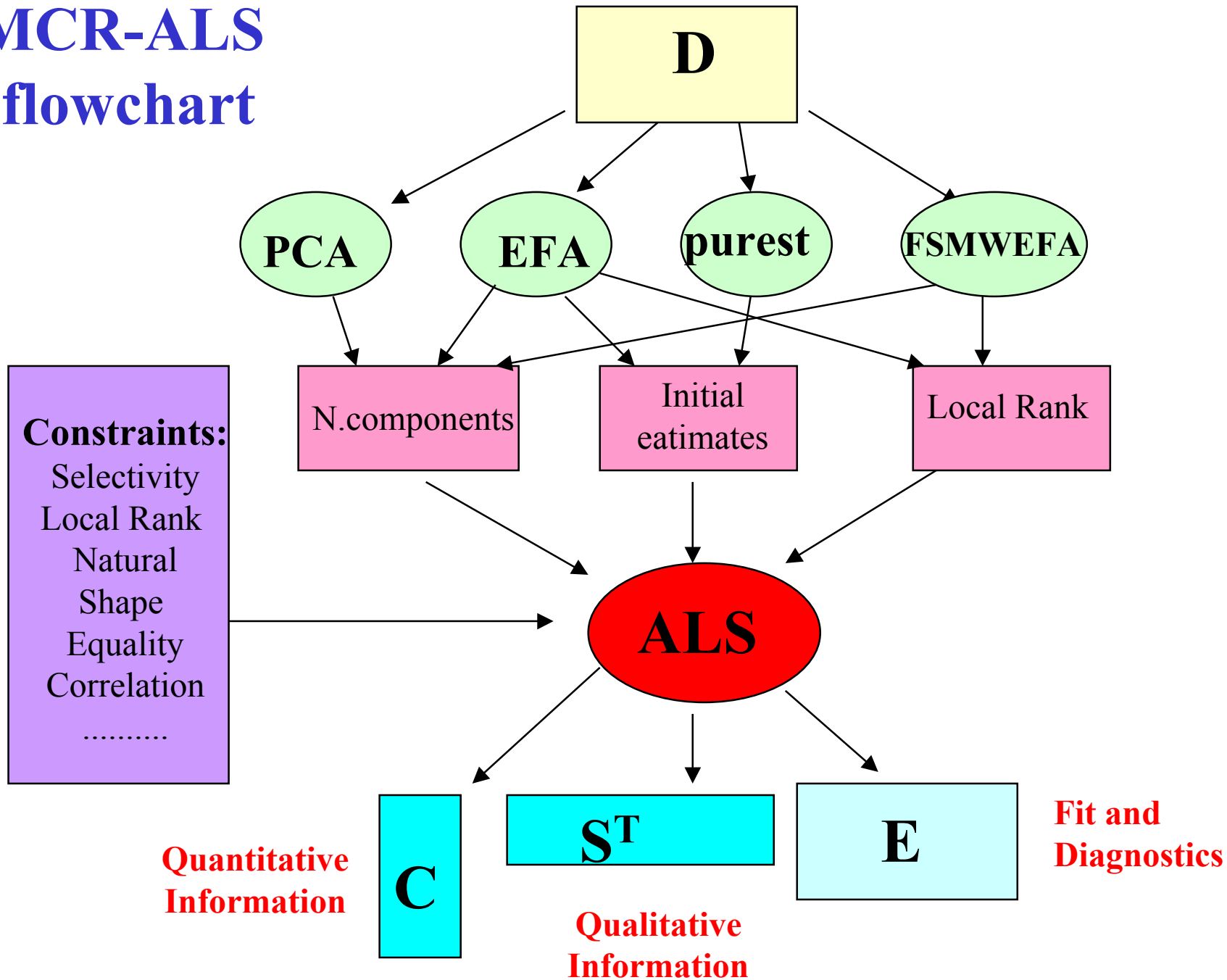
$$1) \min_{\mathbf{S}^T \text{ constrain}} \left\| \mathbf{D} - \mathbf{C} \mathbf{S}^T \right\|$$

$$\mathbf{C}^+ \mathbf{D} = (\mathbf{S}^T)_{uncons} \underset{constrain}{\Rightarrow} \mathbf{S}^T$$

$$2) \min_{\mathbf{C} \text{ constrain}} \left\| \mathbf{D} - \mathbf{C} \mathbf{S}^T \right\|$$

$$\mathbf{D} (\mathbf{S}^T)^+ = (\mathbf{C})_{uncons} \underset{constrain}{\Rightarrow} \mathbf{C}$$

# MCR-ALS flowchart



# Quantitative Information from MCR and Scale (intensity) Ambiguities:

$$d_{ij} = \sum_n c_{in} s_{nj} = \sum_n k c_{in} \frac{1}{k} s_{nj}$$

***k is arbitrary. How to find the right one?***

- Once rotational ambiguities are solved for the species of interest (analyte), how to break intensity ambiguities?
- Is it possible to recover quantitative information using MCR-ALS?



# Solving intensity ambiguities in MCR-ALS

## **Two-way data:**

**In the analysis of a single data matrix intensity-scale ambiguities can be solved using:**

- a) scale/normalization/closure constraints**
- b) external knowledge and equality/correlation constraints**

## **Three-way data**

**In the simultaneous analysis of multiple data matrices intensity/scale ambiguities can be solved**

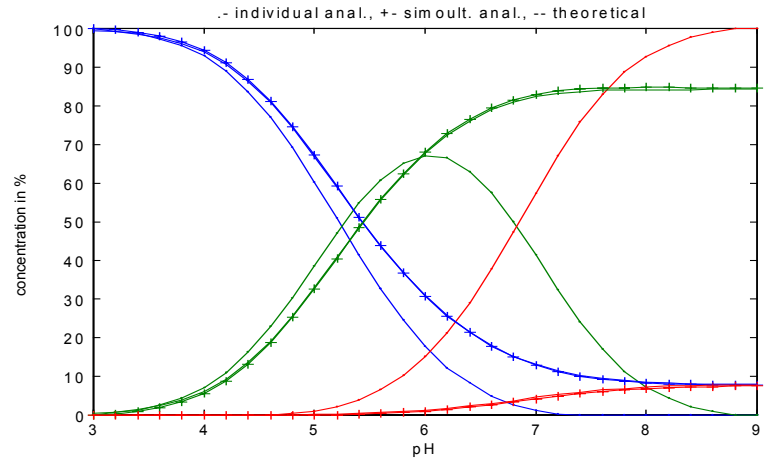
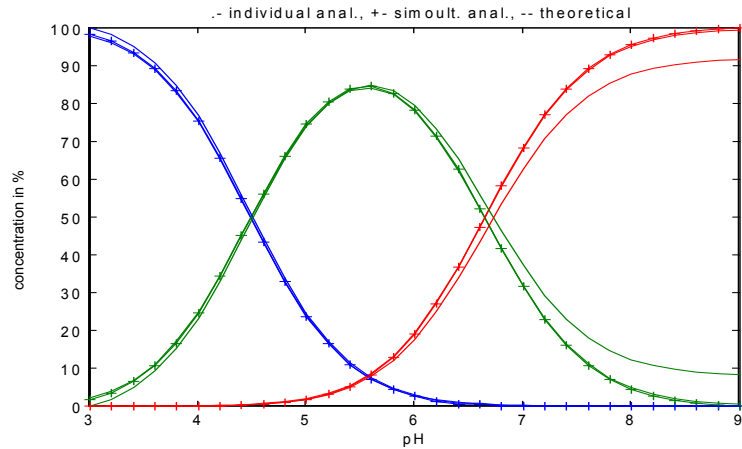
- a) in relative terms (directly)**
- b) in absolute terms using external knowledge**

# Outline

- Introduction to quantitative MCR-ALS method
- **Quantitative MCR-ALS for two-way data**
- Quantitative MCR-ALS for multiway data
- Conclusions and Acknowledgements

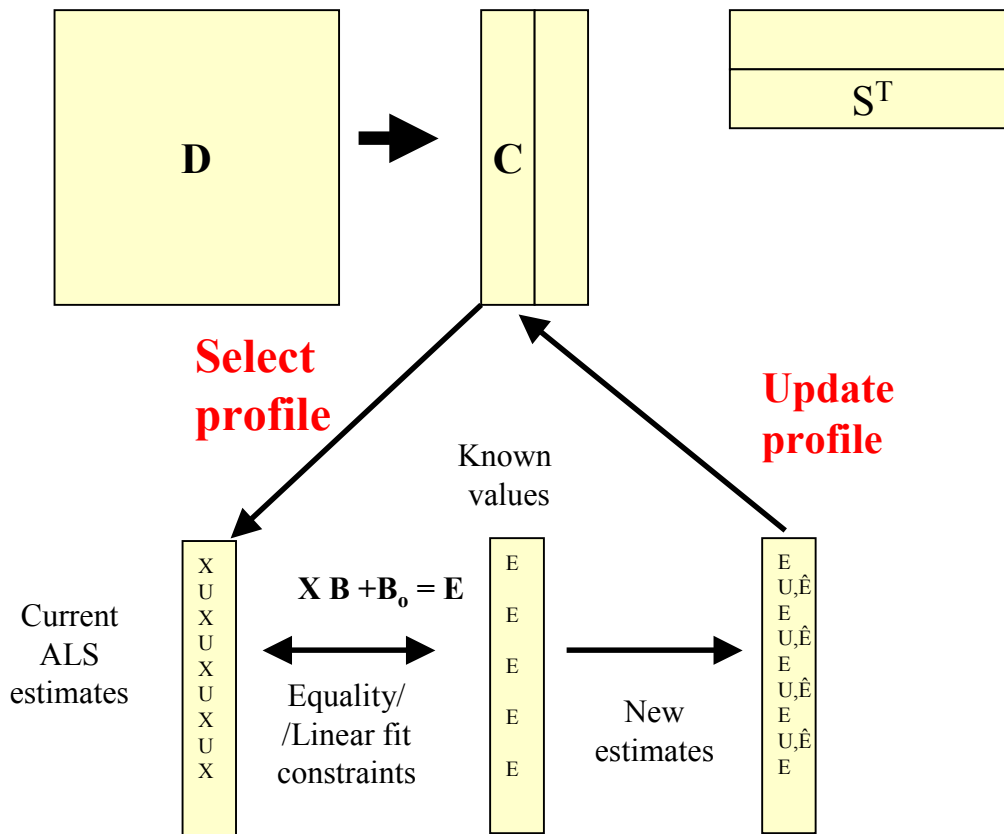
# Solving intensity ambiguities using normalization/closure constraints

**When rotational ambiguities are totally solved, closure constraints give the correct quantitative relationships between the components involved in the closure constraint**



# Solving intensity ambiguities using equality/ correlation constraints

## ALS decomposition

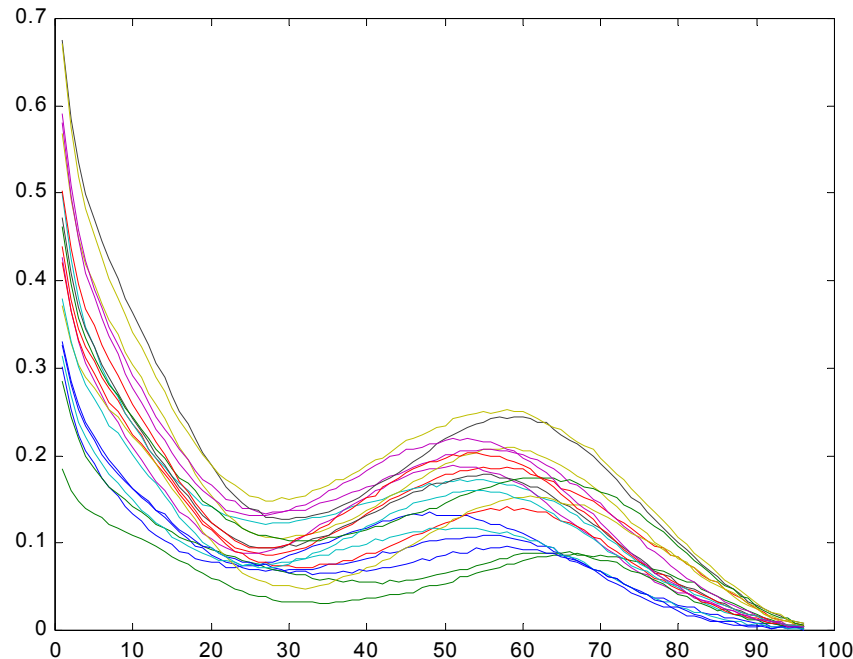


During the ALS decomposition, concentration profiles may be updated using equality constraints or accordingly to their correlation with previously known values ( $E$ )

**Equality constraints:** update  $X$  values by corresponding known  $E$  values

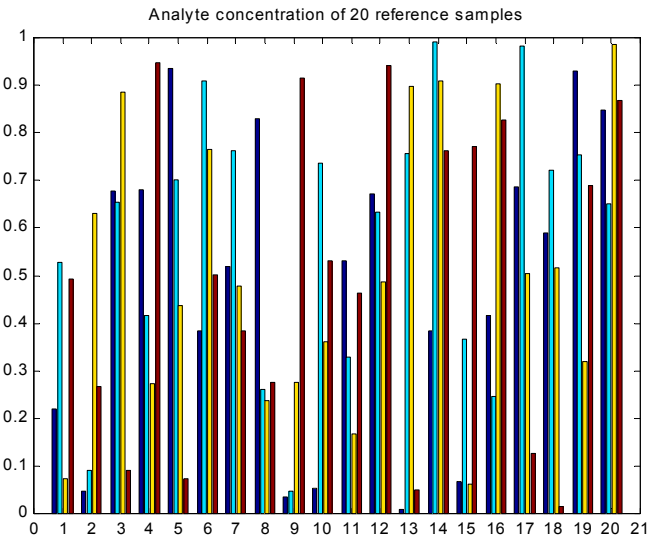
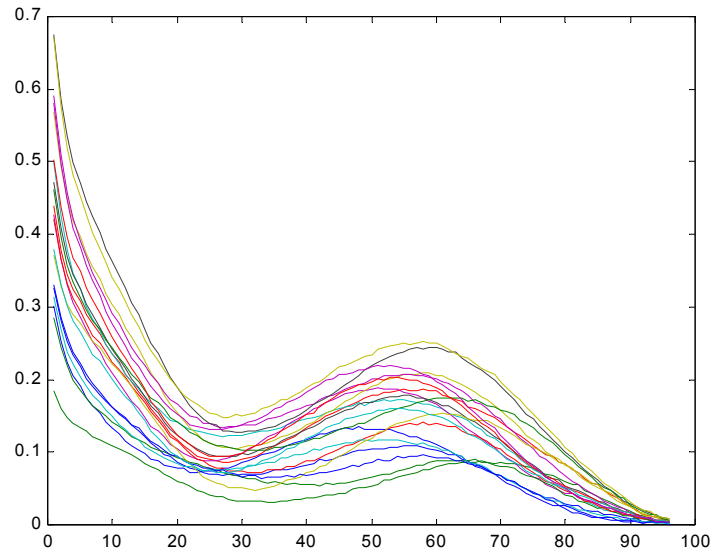
**Correlation constraints:** update  $X$  and  $U$  values by calculated  $\hat{E}$  values deduced from correlation with known  $K$  values,  $X B + B_0 = E$ ;  $U B + B_0 = \hat{E}$

# Example of application of equality constraints to two-way data (simulated data study)



*Problem to solve: predict the concentration of one nucleic base in a mixture of four of them (one analyte in the presence of three interferences)*

# Building the calibration data set

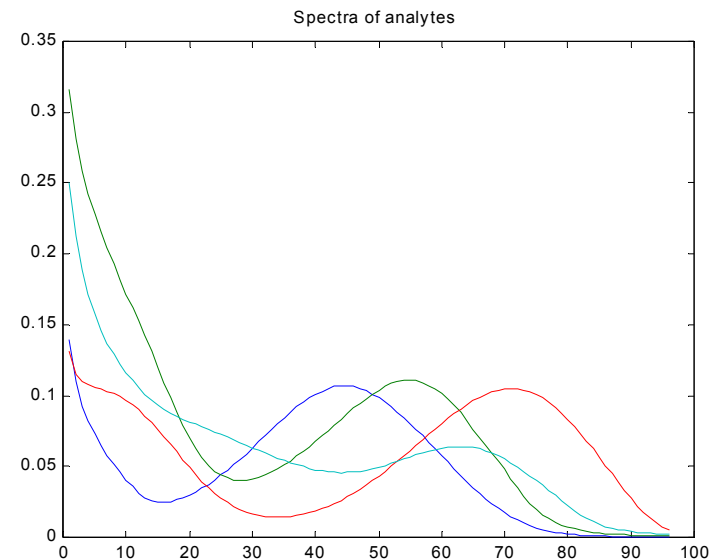


**20 calibration samples with random concentrations of 4 bases between 0-1**

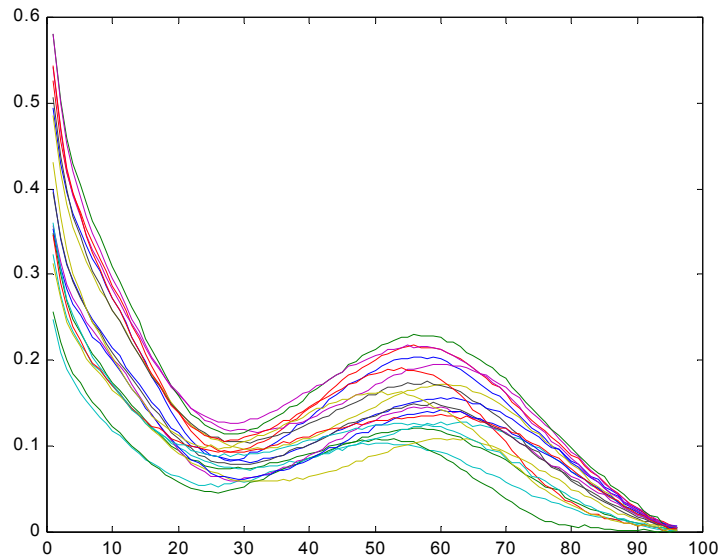
lof (%) = 0.6458

$R^2 = 0.99996$

sd white noise 0.001 units



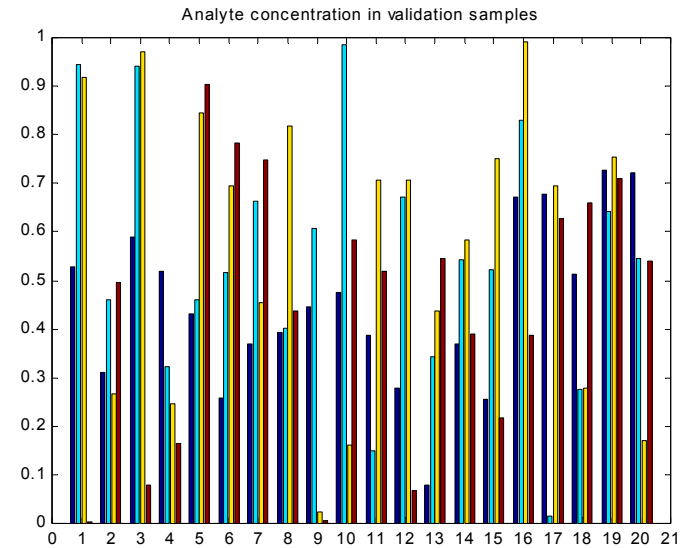
# Building the validation data set



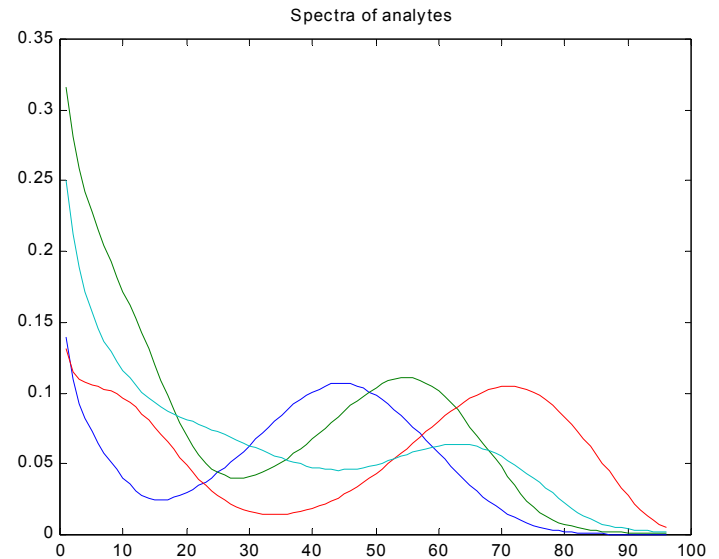
lof (%) = 0.679

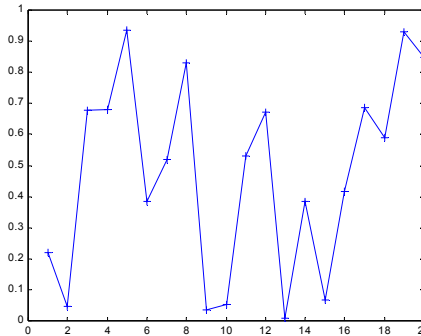
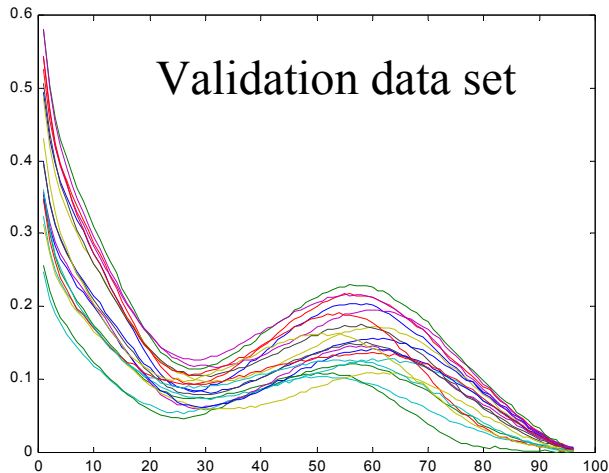
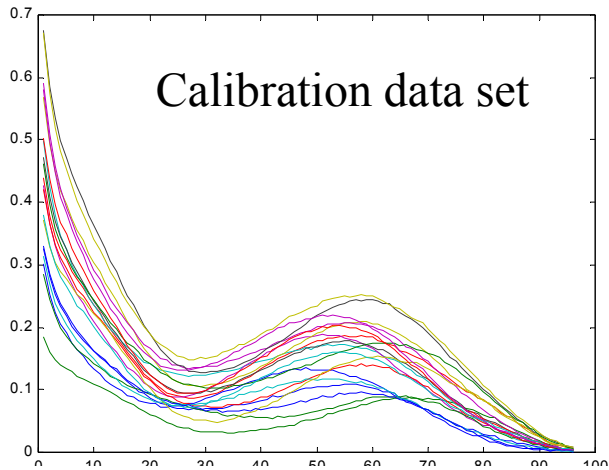
$R^2 = 0.99995$

sd white noise 0.001 units



**20 validation samples with random concentrations of 4 bases between 0-1**

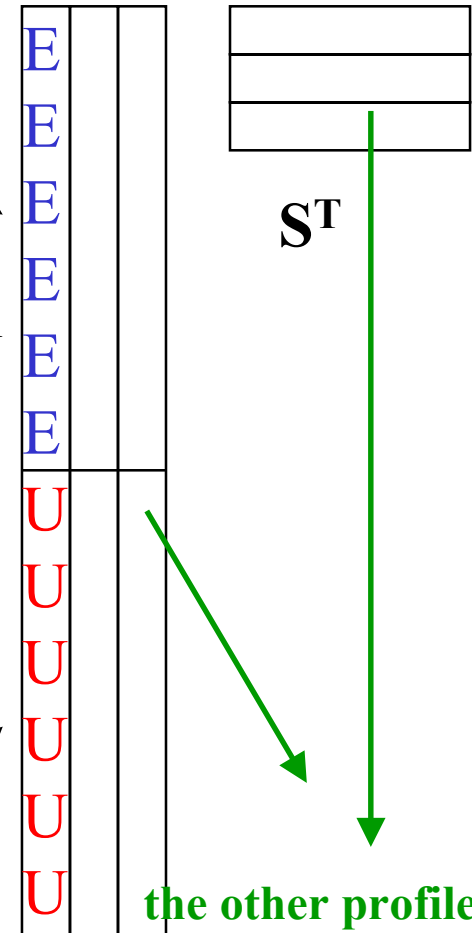




constraints: 1) non-negativity  
2) concentration of the analyte in the calibration set (equality constraint)

**Built a MCR-ALS Model**

**?????**  
**What is the concentration of analyte in the validation data set?**



**C**

**the other profiles are only constrained to be non-negative**



## **Proposed strategy/procedure:**

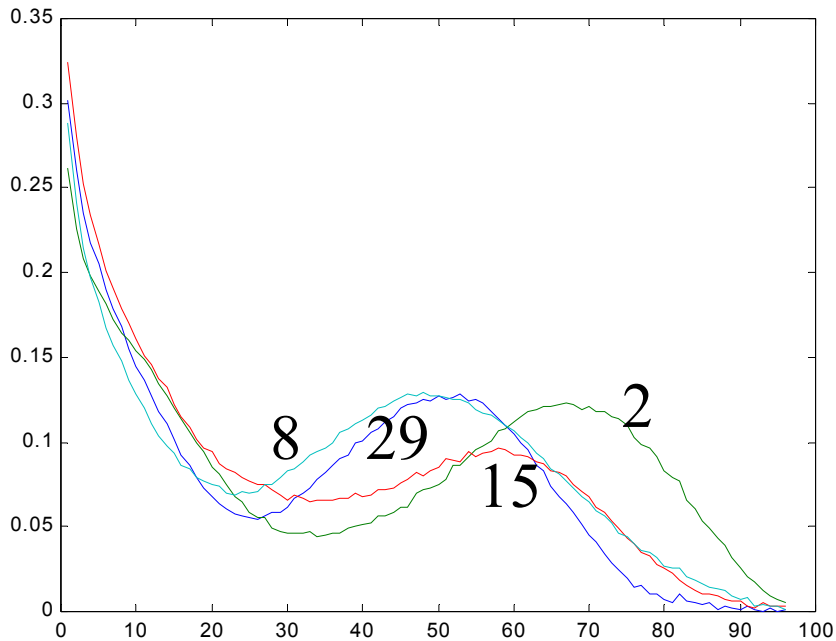
**Apply MCR-ALS using the following constraints:**

- ✓ non-negativity
- ✓ known concentrations of analyte in the calibration data set (equality/correlation constraints)

**Initial estimates for ALS optimization:**

- ✓ from ‘purest’ spectra of the data set (using an approach similar to SIMPLISMA)

# Initial estimations of pure spectra are obtained from the purest samples (key set) number 2, 15, 8 and 29

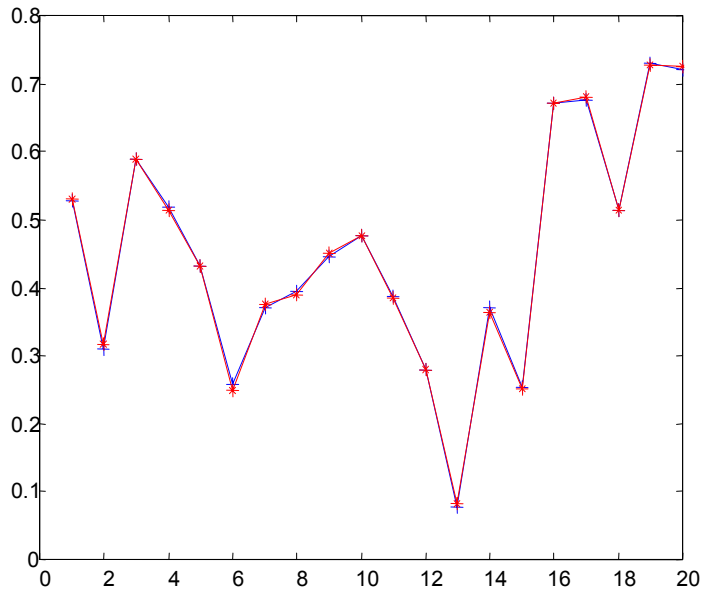


Correlation between true and initially  
estimated spectra  
analyte

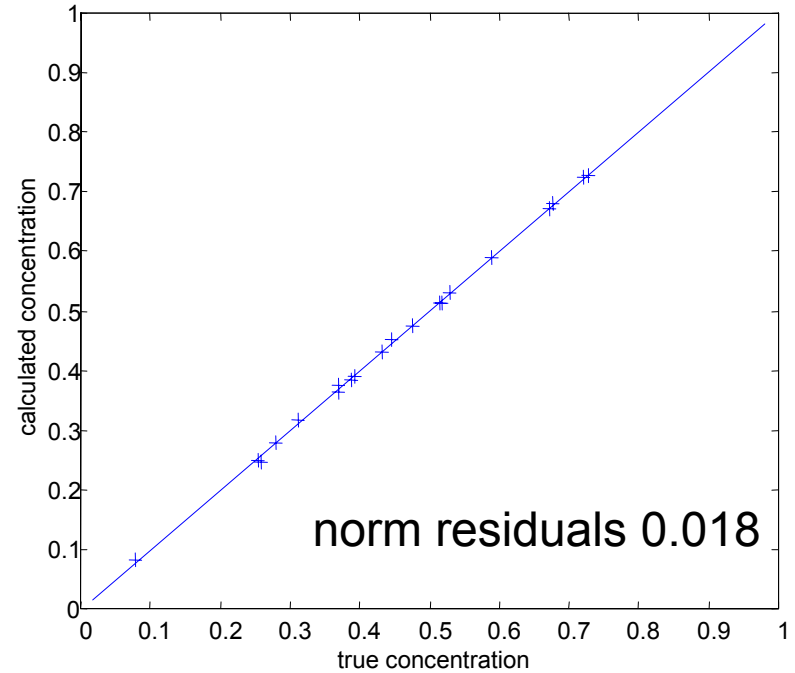
spectrum	1	2	3	4
2	0.0470	0.0920	<b>0.6316</b>	0.2661
8	<b>0.8310</b>	0.2625	0.2378	0.2771
15	0.0668	0.3653	0.0606	<b>0.7702</b>
29	0.4469	<b>0.6056</b>	0.0221	0.0063

# MCR-ALS concentration prediction for **analyte 1**

predicted (red) versus true (blue) concentrations of analyte 1



$$y = -0.0023 + 1.0044x$$



**RECOVERED  
CONCENTRATIONS  
FOR ANALYTE 1  
IN THE VALIDATION  
DATA SET USING  
MCR-ALS AND PLS  
(without any data  
pretreatment)**

	<b>CONCENTRATIONS</b>			<b>% Error</b>		
	<b>ALS</b>	<b>PLS</b>	<b>'true'</b>	<b>ALS</b>	<b>PLS</b>	
1	0.5295	0.5298	0.5285	1	0.1708	0.2405
2	0.3159	0.3160	0.3107	2	1.6500	1.7056
3	0.5882	0.5885	0.5881	3	0.0127	0.0646
4	0.5144	0.5142	0.5181	4	-0.7200	-0.7459
5	0.4317	0.4316	0.4308	5	0.1917	0.1650
6	<b>0.2478</b>	<b>0.2478</b>	<b>0.2588</b>	6	<b>-4.2732</b>	<b>-4.2555</b>
7	0.3759	0.3761	0.3702	7	1.5321	1.5840
8	0.3898	0.3898	0.3930	8	-0.8076	-0.8169
9	0.4505	0.4501	0.4469	9	0.8052	0.7304
10	0.4763	0.4767	0.4759	10	0.0850	0.1647
11	0.3840	0.3838	0.3878	11	-0.9876	-1.0488
12	0.2783	0.2786	0.2793	12	-0.3624	-0.2369
13	<b>0.0818</b>	<b>0.0819</b>	<b>0.0783</b>	13	<b>4.5376</b>	<b>4.5876</b>
14	0.3646	0.3647	0.3697	14	-1.3929	-1.3524
15	0.2501	0.2503	0.2539	15	-1.5048	-1.4455
16	0.6703	0.6704	0.6718	16	-0.2247	-0.2005
17	0.6812	0.6820	0.6762	17	0.7274	0.8485
18	0.5132	0.5129	0.5139	18	-0.1471	-0.1981
19	0.7268	0.7267	0.7286	19	-0.2542	-0.2584
20	0.7247	0.7245	0.7208	20	0.5459	0.5220

# Formulas for Results Validation

RMSEP = Root Mean Square  
Error in Prediction

$$\text{RMSEP} = \sqrt{\frac{\sum_i \sum_j (\hat{c}_{ij} - c_{ij})^2}{\text{NR}}}$$

SEP = Standard Error in  
Prediction (bias corrected)

$$\text{SEP} = \sqrt{\frac{\sum_i \sum_j (\hat{c}_{ij} - c_{ij} - \text{bias})^2}{\text{NR} - 1}}$$

bias = systematic deviations

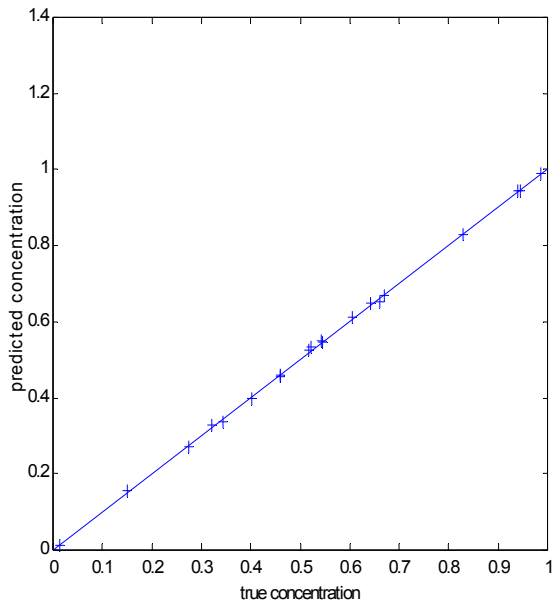
$$\text{bias} = \sqrt{\frac{\sum_i \sum_j (\hat{c}_{ij} - c_{ij})}{\text{NR}}}$$

RE = Overall % relative error  
in the prediction

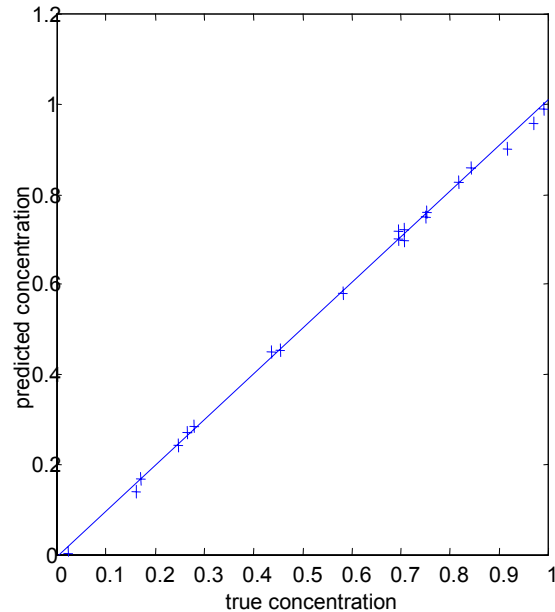
$$\text{RE} = 100 \sqrt{\frac{\sum_i \sum_j (\hat{c}_{ij} - c_{ij})^2}{\sum_i \sum_j (c_{ij})^2}}$$

# MCR-ALS concentration prediction for other analytes

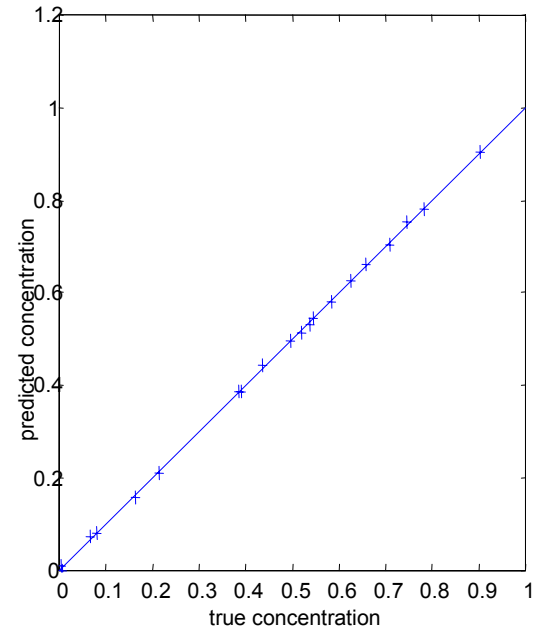
## Analyte 2



## Analyte 3

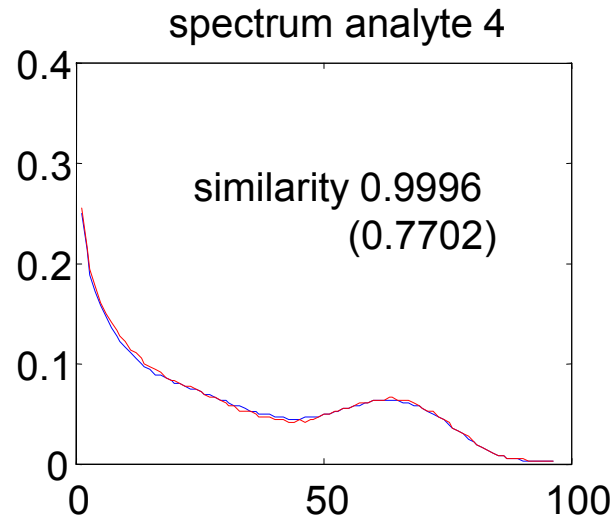
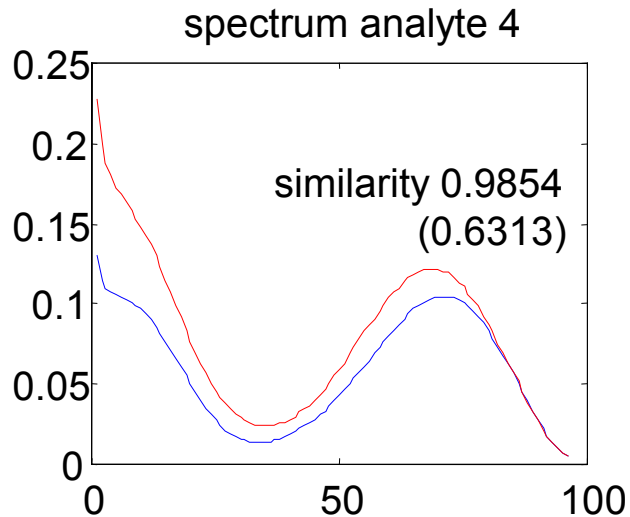
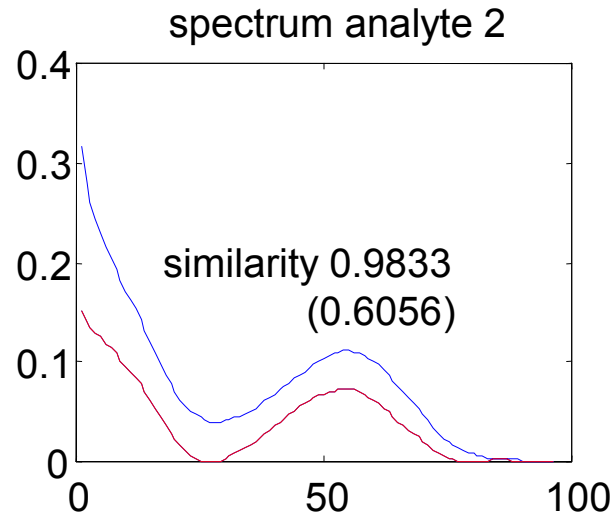
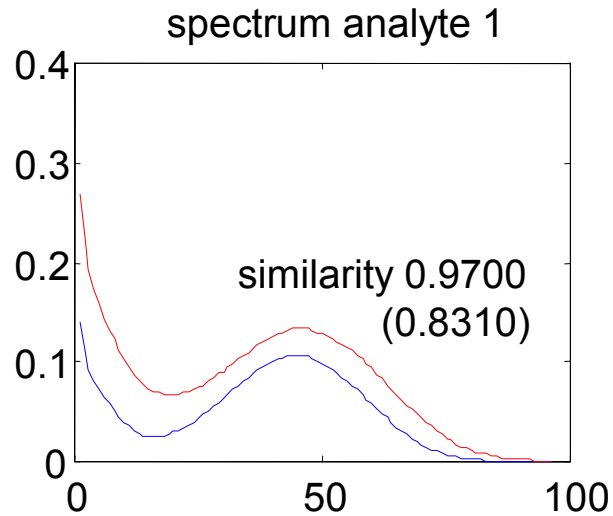


## Analyte 4



analyte	RMSEP	SEP	bias	RE %	slope	offset	corcoef	sdres
1.0000	0.0041	0.0042	0.0003	1.7714	1.0044	-0.0023	0.9997	0.0179
2.0000	0.0050	0.0050	-0.0012	1.4081	1.0019	0.0002	0.9998	0.0217
3.0000	0.0117	0.0120	-0.0001	2.8427	1.0092	-0.0052	0.9992	0.0511
4.0000	0.0046	0.0047	0.0007	1.7384	1.0010	-0.0011	0.9999	0.0205

# MCR-ALS spectra prediction



In parenthesis correlation (similarity) respect initial estimates  
red predicted, blue 'true'

## Example of application (real data study):

Quantitation of forage samples using NIR spectroscopy and MCR-ALS with known concentration constraints

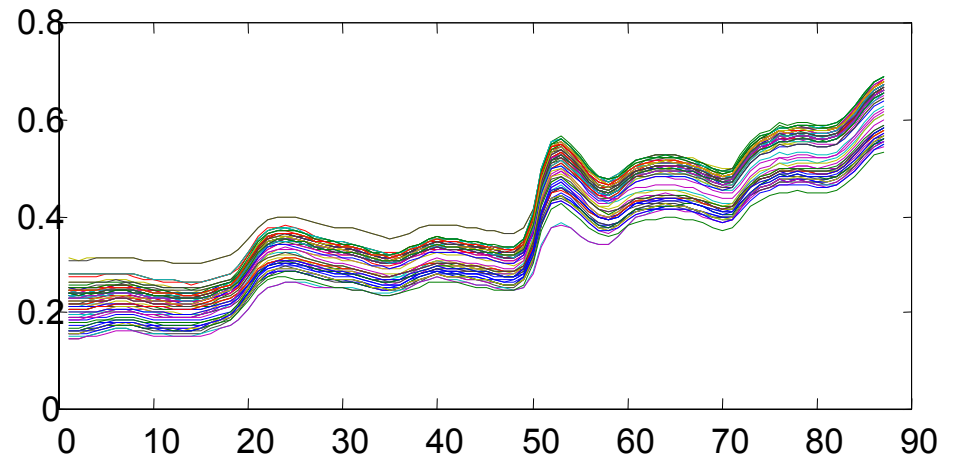
Problem to solve: Determination of moisture and protein content using a calibration and a validation real data sets

Comparison with results obtained using PLS multivariate calibration

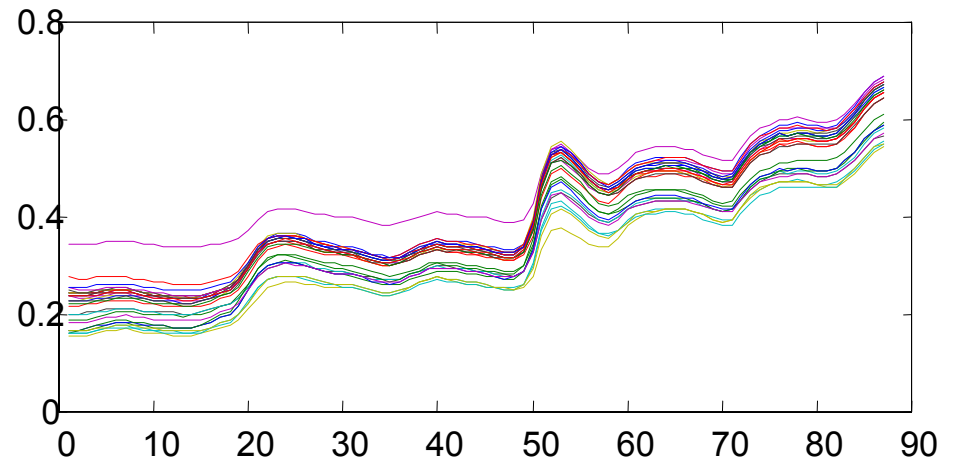


# Example of application: NIR real data

Calibration data set  
86 samples  
87 wavelengths  
no-pretreatment

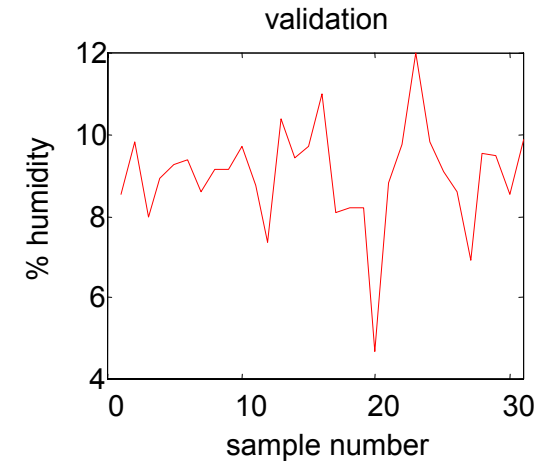
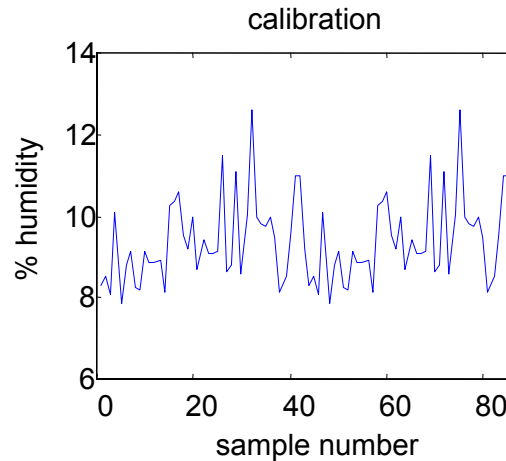


Validation data set  
31 samples  
87 wavelengths  
no-pretreatment

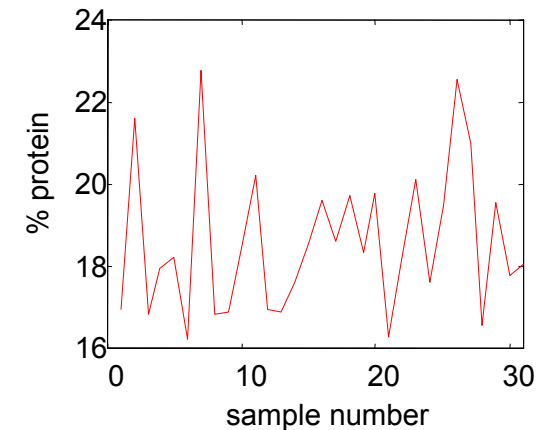
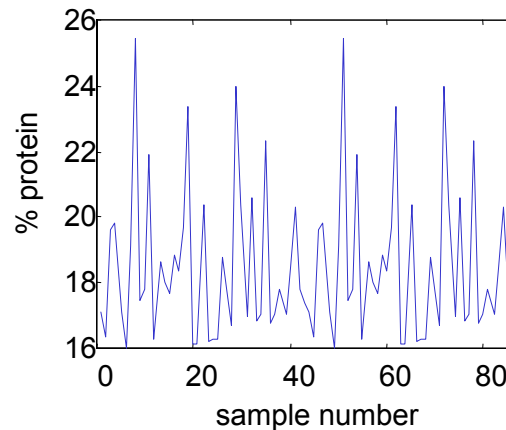


# Determination of moisture and protein

Moisture concentrations  
in the calibration and  
validation data sets



Protein concentrations  
in the calibration and  
validation data sets



# NIR Data structure:

*Singular Values*

*PCA lof*

**Raw data**

**Detection of purest spectra**

**39.8310**

**2.68%**

**0.9829**

**1.06%**

**0.3629**

**0.55%**

**0.1889**

**0.28%**

0.0736

0.21%

0.0562

0.15%

0.0334

0.13%

0.0314

0.10%

0.0227

0.09%

0.0205

0.07%

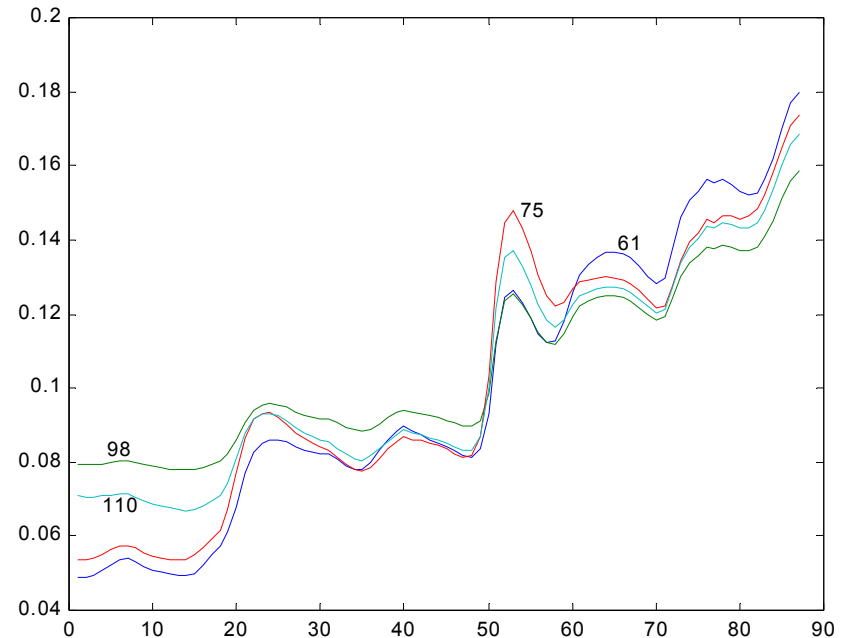
num	moist	protein
-----	-------	---------

61	9.51	19.7
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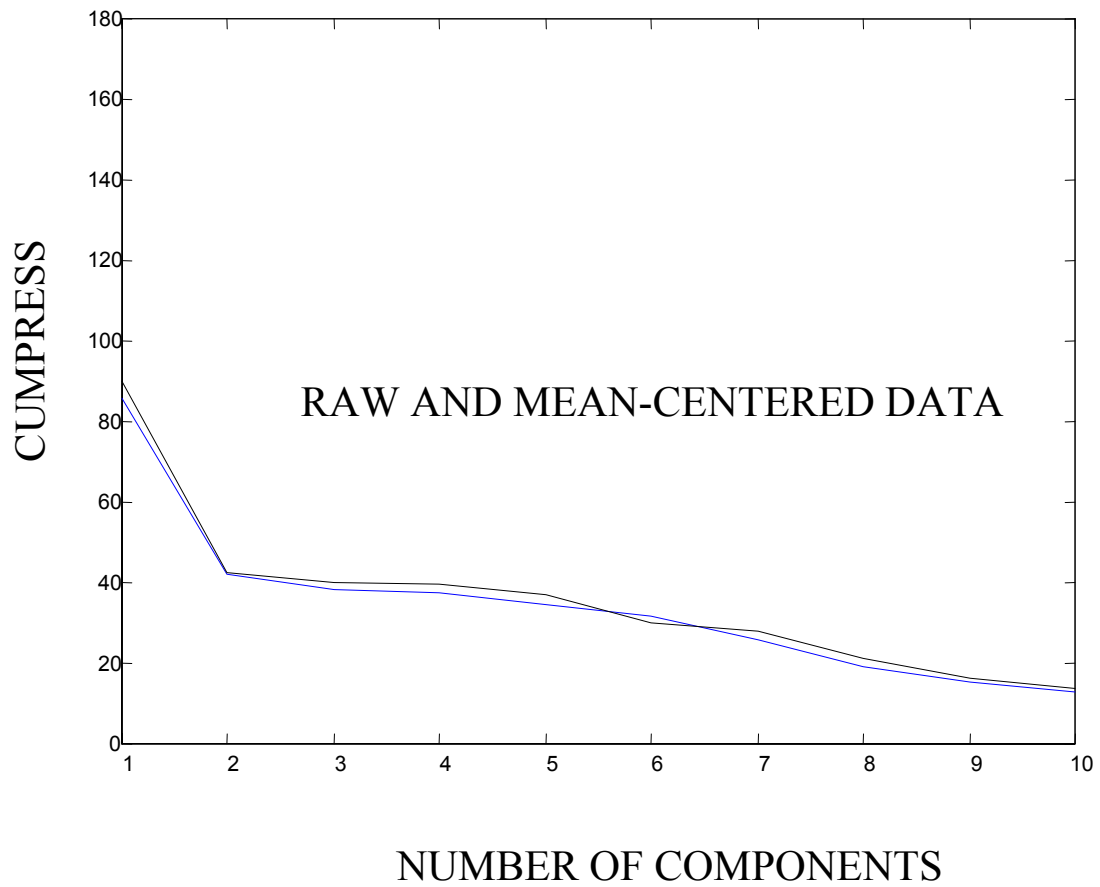
98	7.33	16.93
----	------	-------

<b>75</b>	<b>12.6</b>	<b>20.58</b>
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110	9.80	17.60
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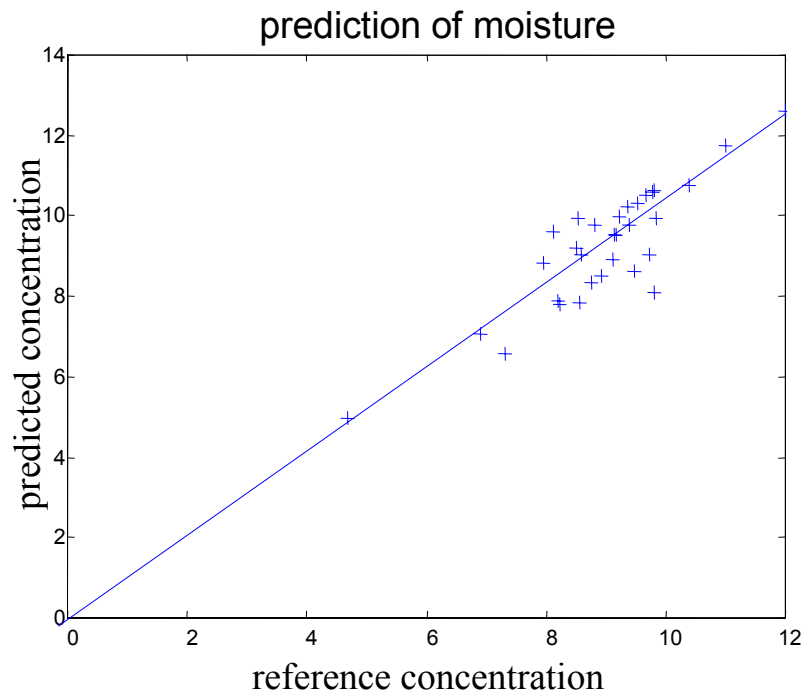


# PLS CROSS-VALIDATION MOISTURE

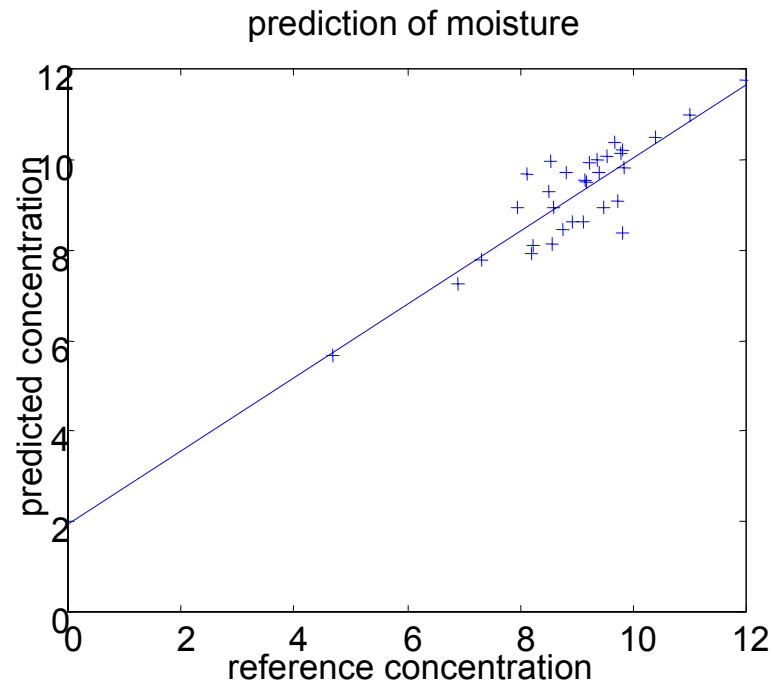


# Prediction of moisture (raw data)

## ALS predictions



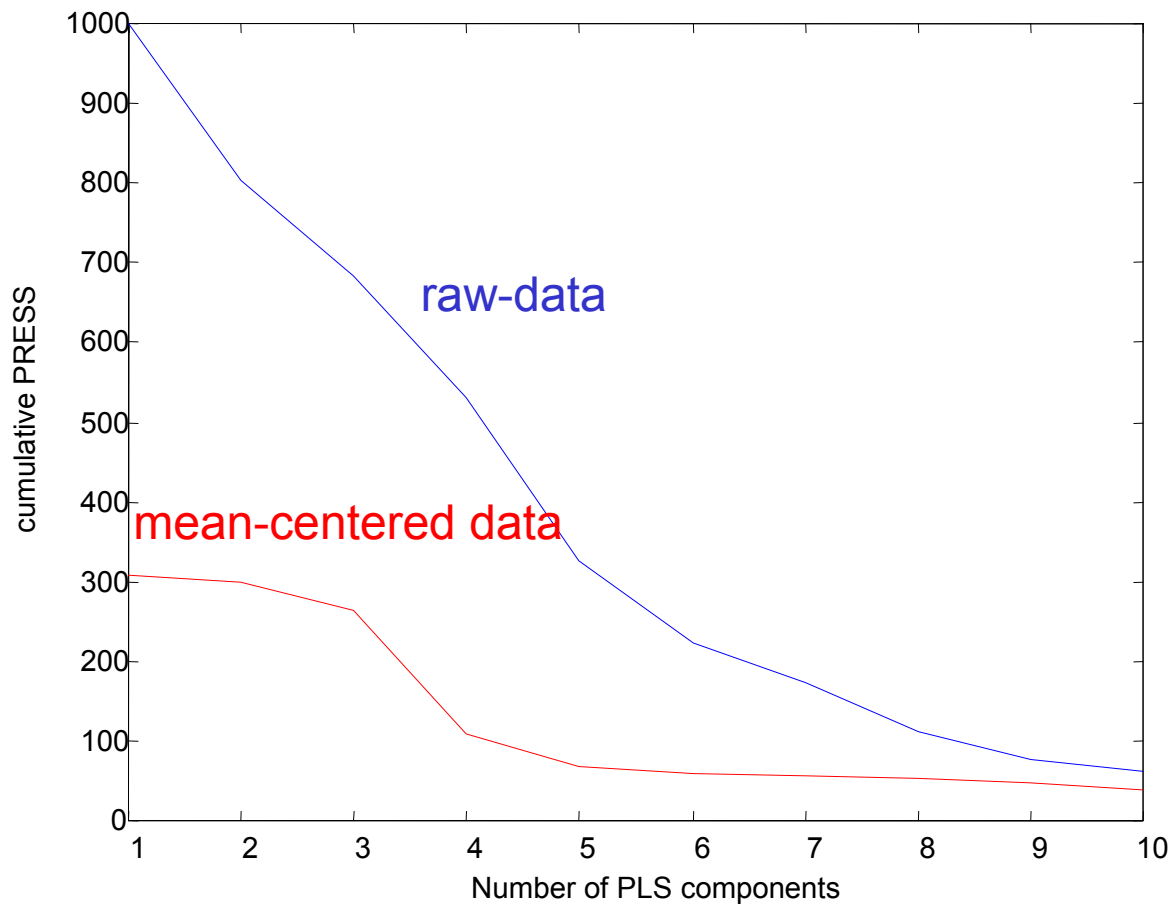
## PLS predictions



method	analyte	RMSEP	SEP	bias	RE %	slope	offset	corcoef	sdres
ALS(4)	moisture	0.7470	0.7227	-0.2291	0.9071	1.0531	-0.2478	0.8827	3.94
PLS(4)	moisture	0.6760	0.6433	-0.2375	0.8208	0.8097	1.9475	0.8677	3.26
PLS(3)	moisture	0.6771	0.6621	-0.1849	0.8222	0.8569	1.4711	0.8656	3.48

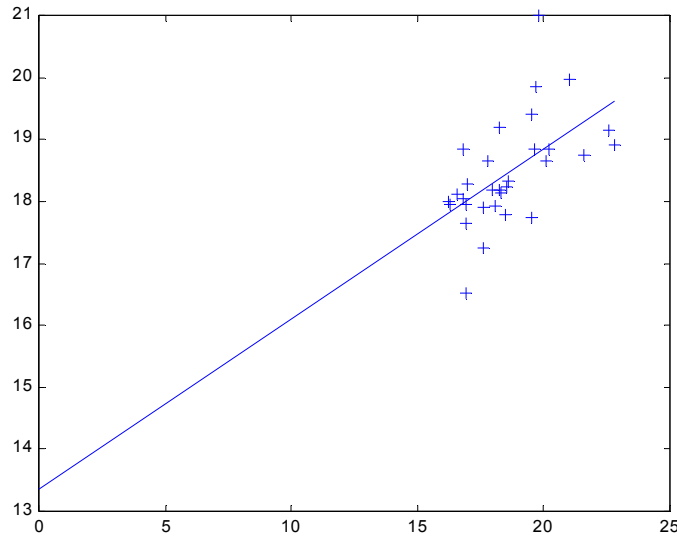
**Results did not improve significantly with mean centering nor with 2<sup>nd</sup> derivative!**

# PLS CROSS-VALIDATION PROTEIN

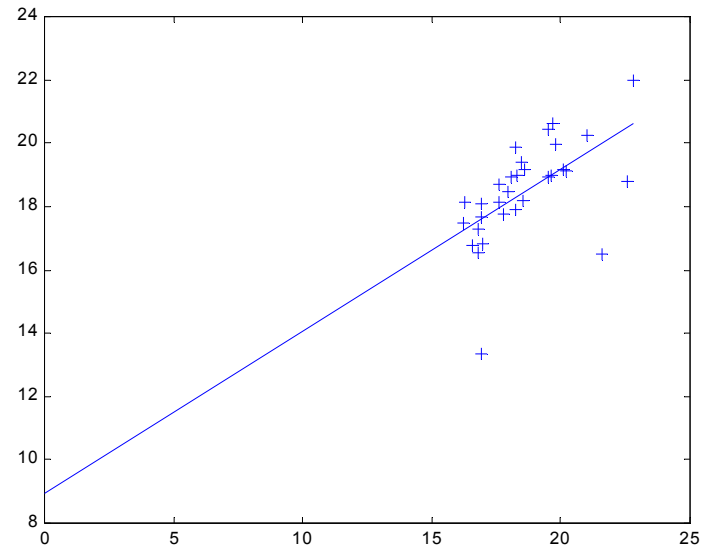


# Prediction of protein (mean centered data)

## ALS 4 components



## PLS 4 components



method	analyte	RMSEP	SEP	bias	RE %	slope	offset	corcoef	sdres
ALS(4)	protein	1.4548	1.4732	0.1259	0.4174	0.2743	13.363	0.5673	3.881
PLS(4)	protein	1.5388	1.5590	0.1259	0.4415	0.5116	8.951	0.5754	7.088
PLS(8)	protein	0.9865	0.9946	0.1259	0.2830	0.7033	5.388	0.8295	4.616
PLS(6)	protein	0.8572	0.8620	0.1259	0.2459	0.7006	5.438	0.8787	3.711
2nd der.									

# Provisory conclusions on the use of MCR-ALS for quantitative two-way data analysis

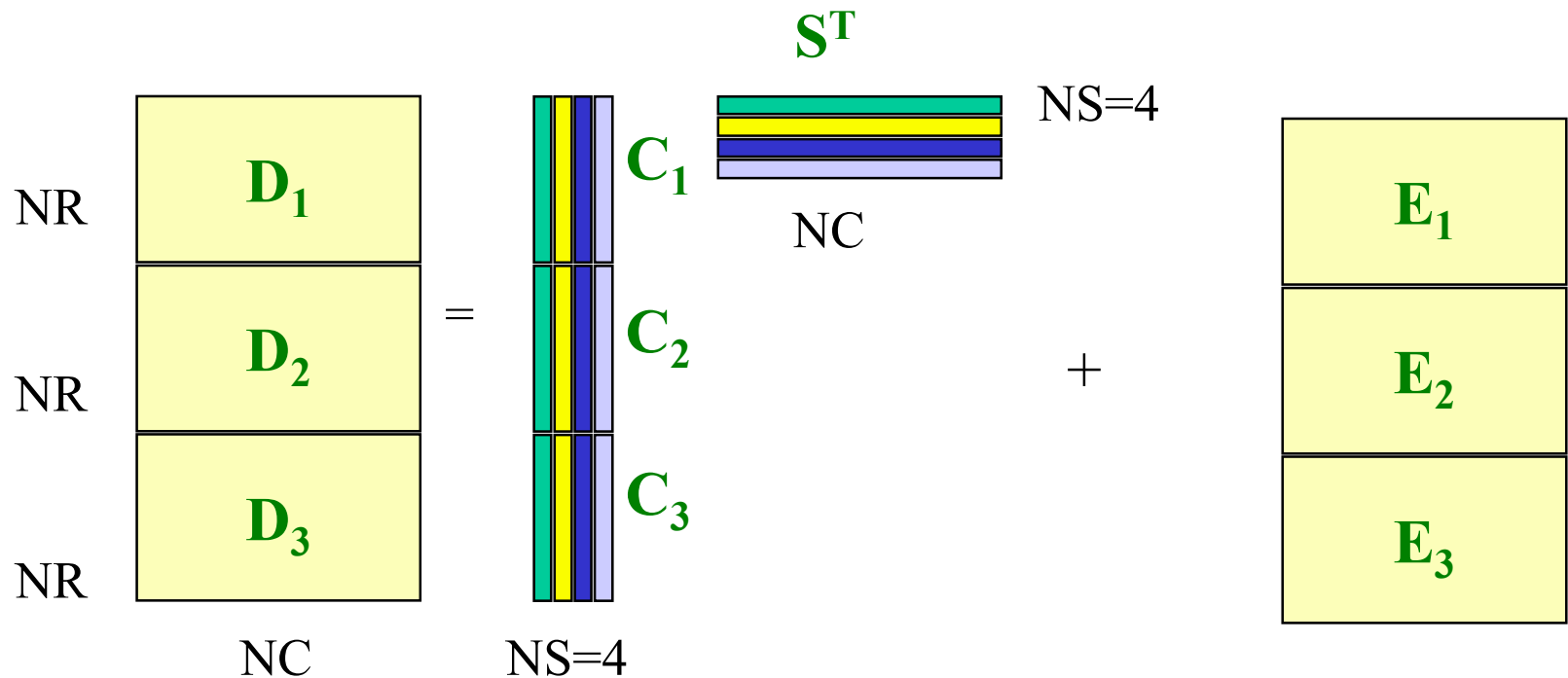
- ✓ **MCR-ALS can be used for the resolution and quantitation of those components which contribute significantly to the signal (i.e. usually for the major constituents of mixtures)**
- ✓ **However, it is not recommended for**
  - **components contributing little to the signal**
  - **when the previous knowledge of the system is minimal (natural samples)**
  - **when nearly no constraints can be applied (i.e. for mean centered 2nd derivative data)**



# Outline

- Introduction to MCR-ALS method
- Quantitative MCR-ALS for two-way data
- **Quantitative MCR-ALS for three-way data**
- Conclusions and Acknowledgements

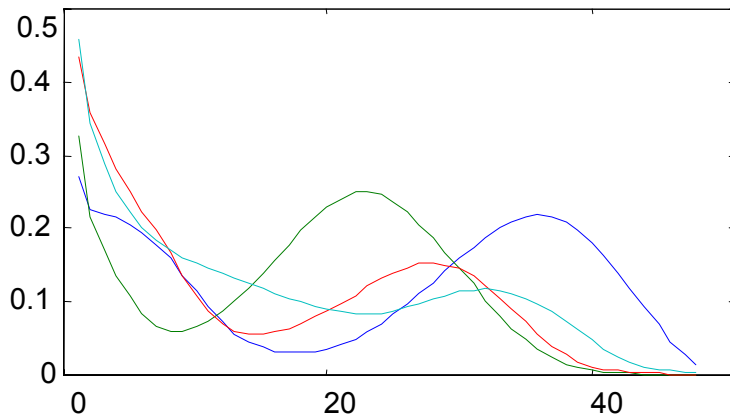
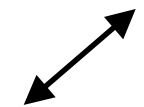
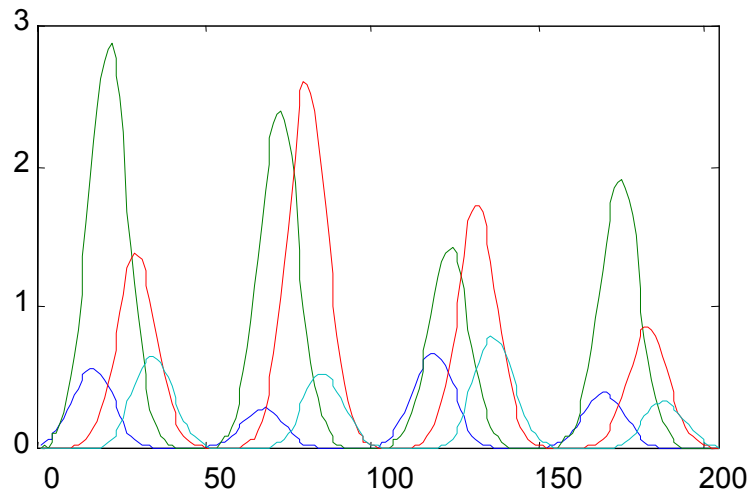
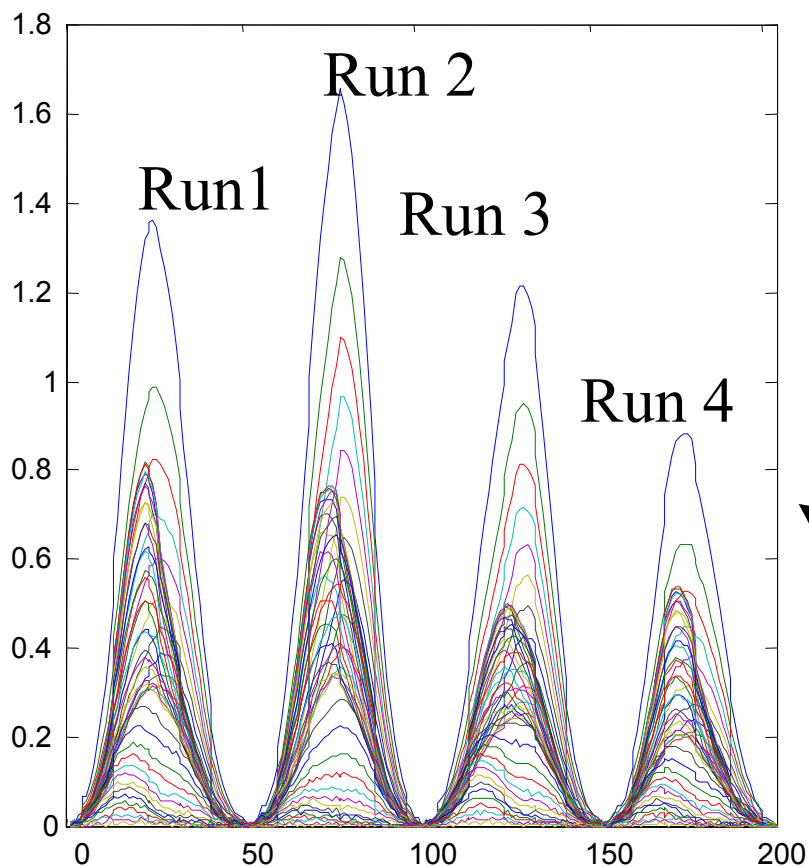
# MCR-ALS can be easily extended to three-way (bilinear) data!



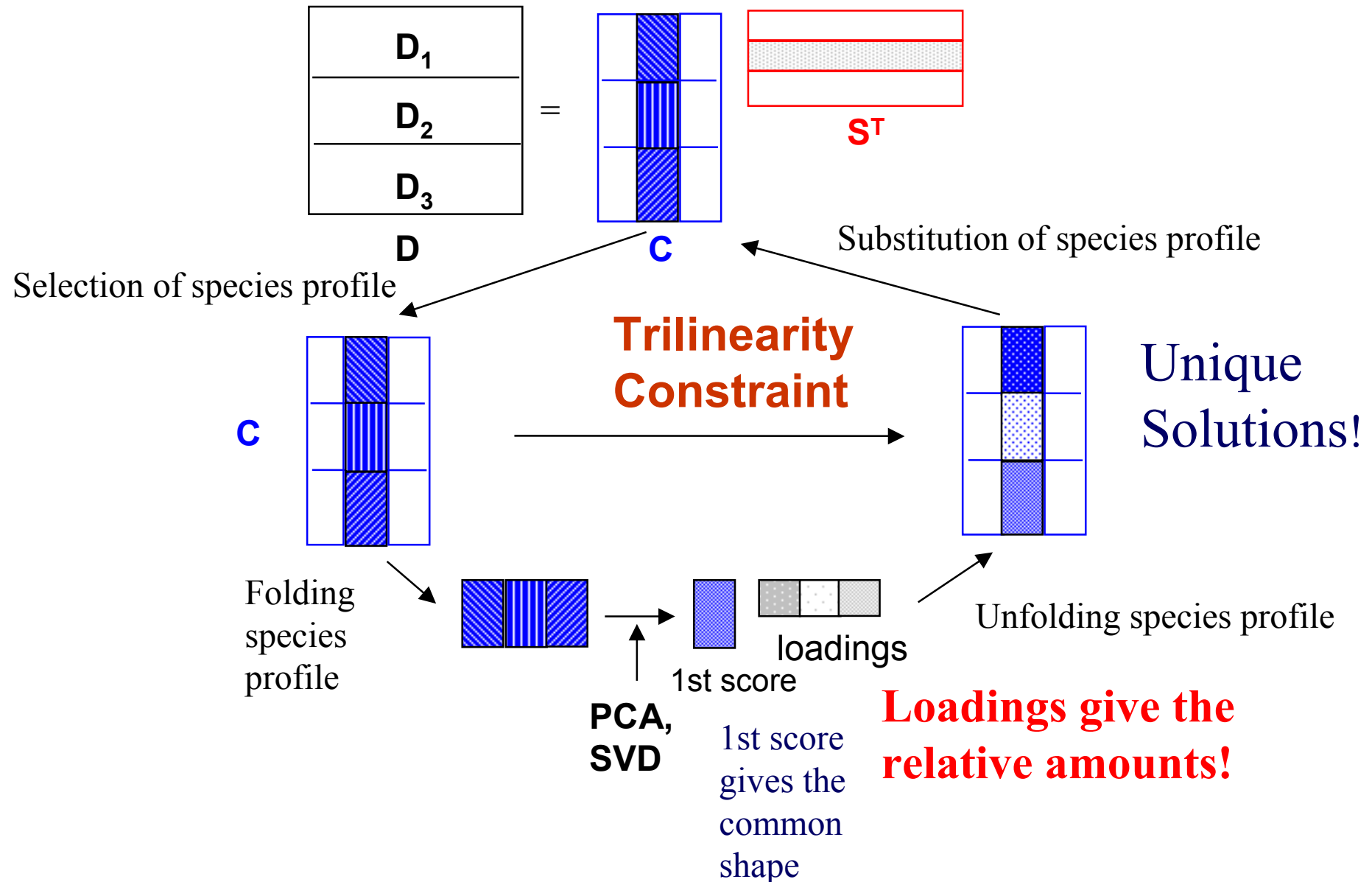
**Column-wise Data Matrix Augmentation constraints may be applied independently to each concentration species profile in each submatrix  $C_1$ ,  $C_2$ ,  $C_3$**

# Trilinear data

$$d_{ijk} = \sum_{n=1}^N c_{in} s_{jn} t_{kn} + e_{ijk}$$

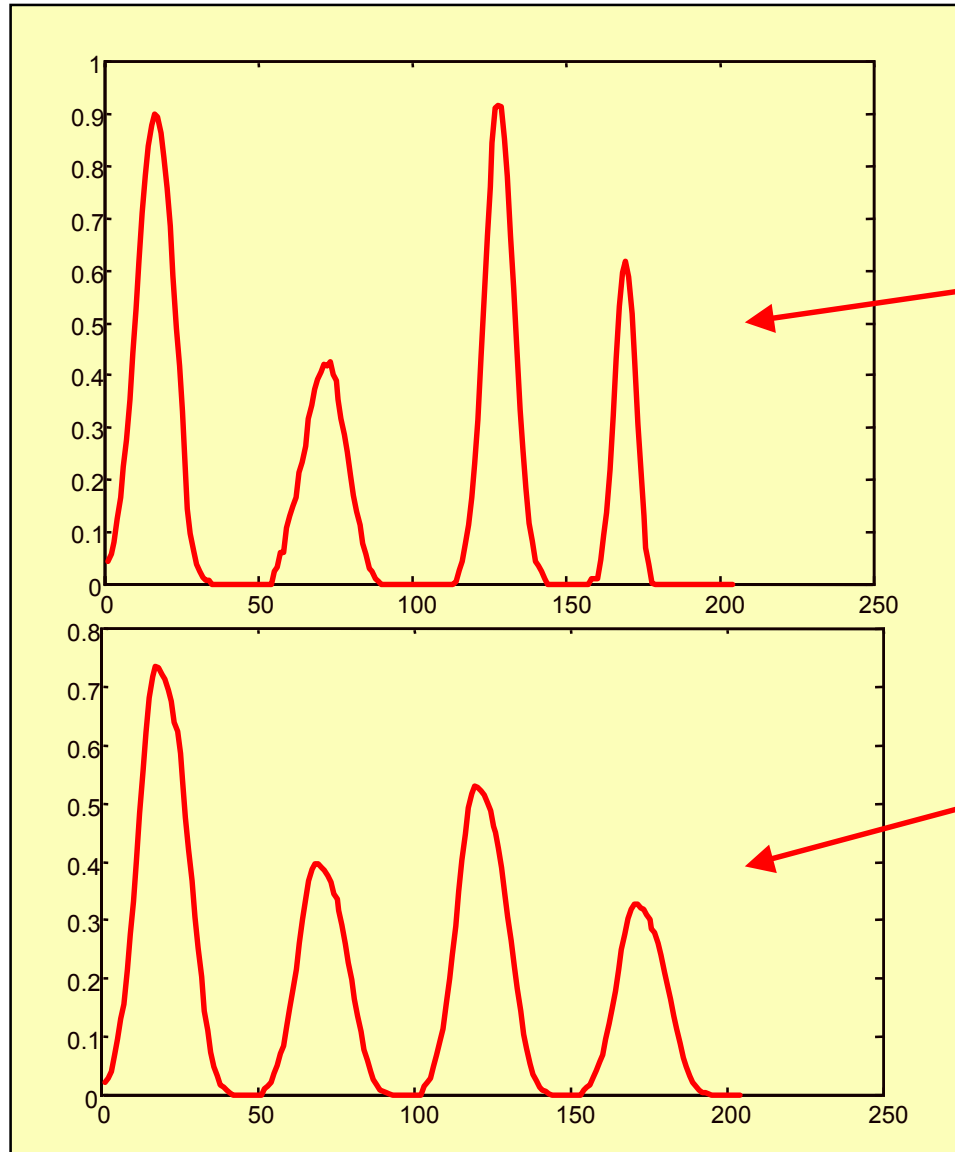


# Trilinearity can be implemented independently for each component (chemical species) in MCR-ALS!



# Effect of application of a trilinearity constraint

Trilinearity  
constraint

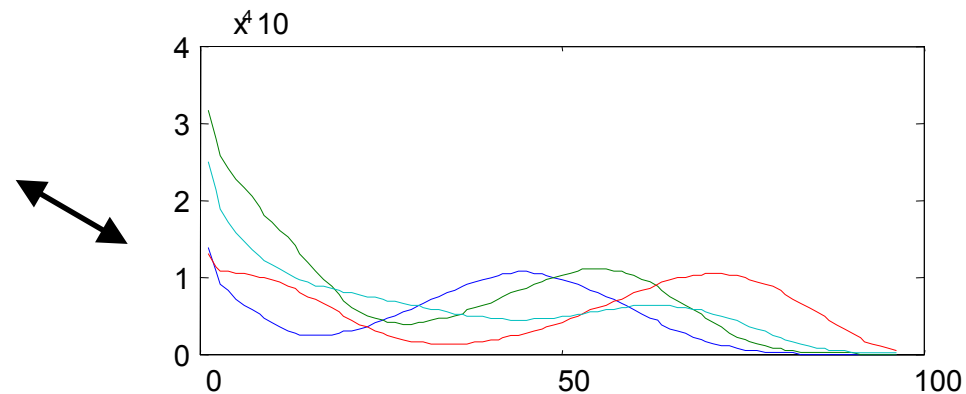
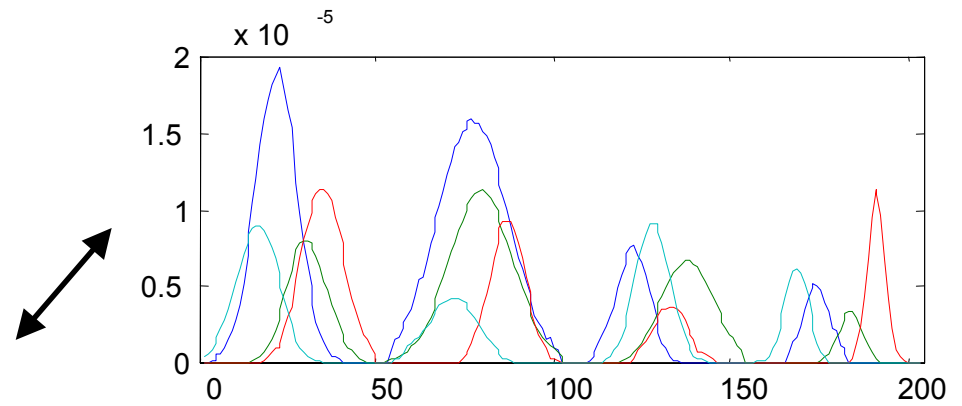
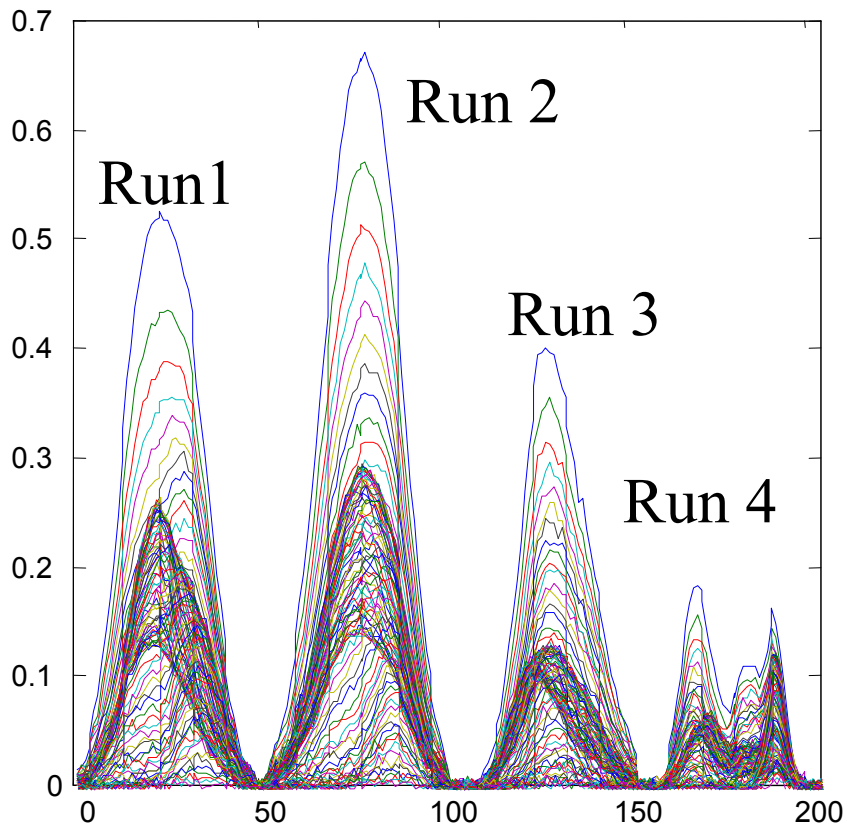


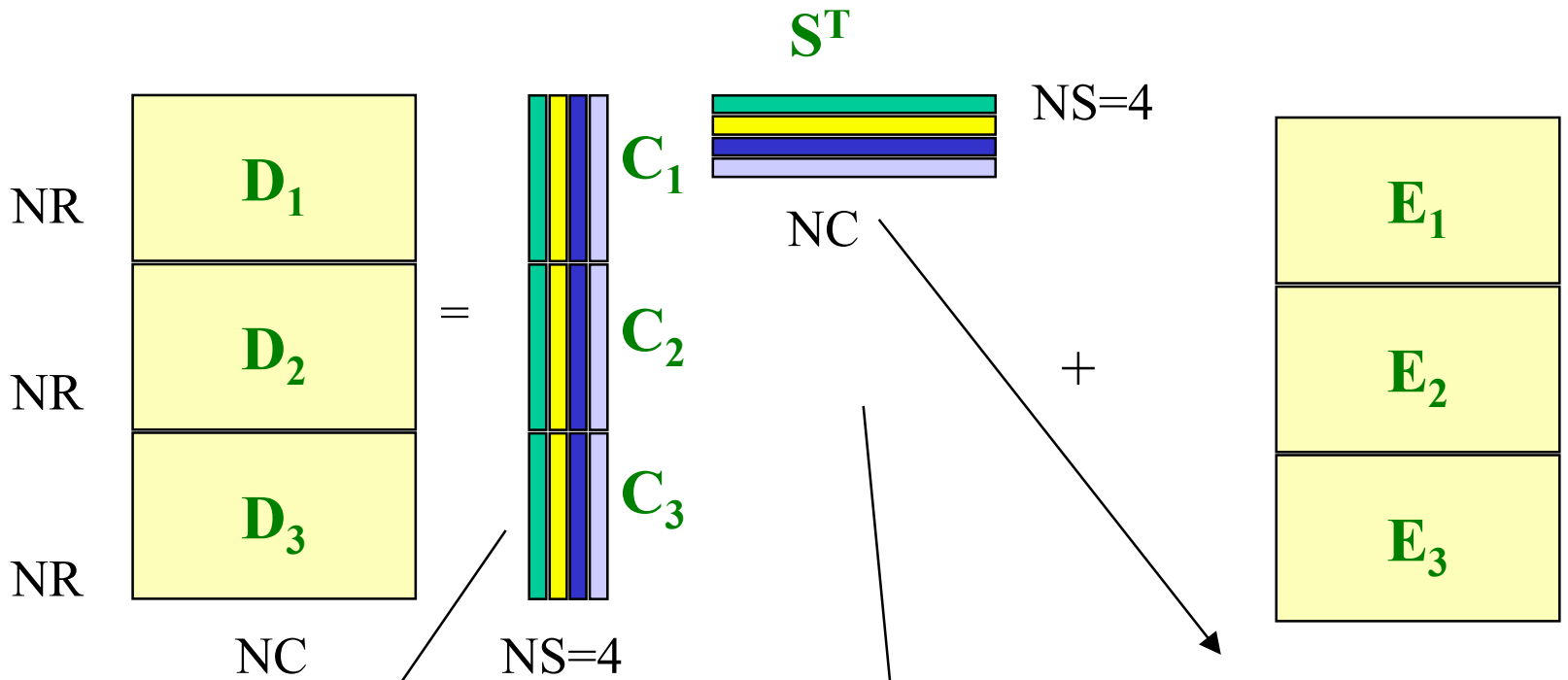
# Trilinear data: spectra recovery

species	TLD (cos)	ALS (cos)	TLD (sin)	ALS (sin)
1	0,9995	0,9999	0,033	0,0107
2	1	1	0,0069	0,0068
3	0,9998	0,9999	0,0221	0,0136
4	0,9999	1	0,0124	0,0086

# Non-trilinear data

$$d_{ijk} \neq \sum_{n=1}^N c_{in} s_{jn} t_{kn} + e_{ijk}; \quad d_{ijk} = \sum_{p=1}^{N_p} \sum_{q=1}^{N_q} \sum_{r=1}^{N_r} g_{pqr} c_{ip} s_{jq} t_{kr} + e_{ijk}$$





**When selectivity or local rank resolution conditions are fulfilled for the concentration profile of one species in one matrix, resolution will be also achieved for the concentration profiles of the same species in the other matrices simultaneously analyzed**

**The set of possible LS solutions for  $S^T$  are now more restricted**

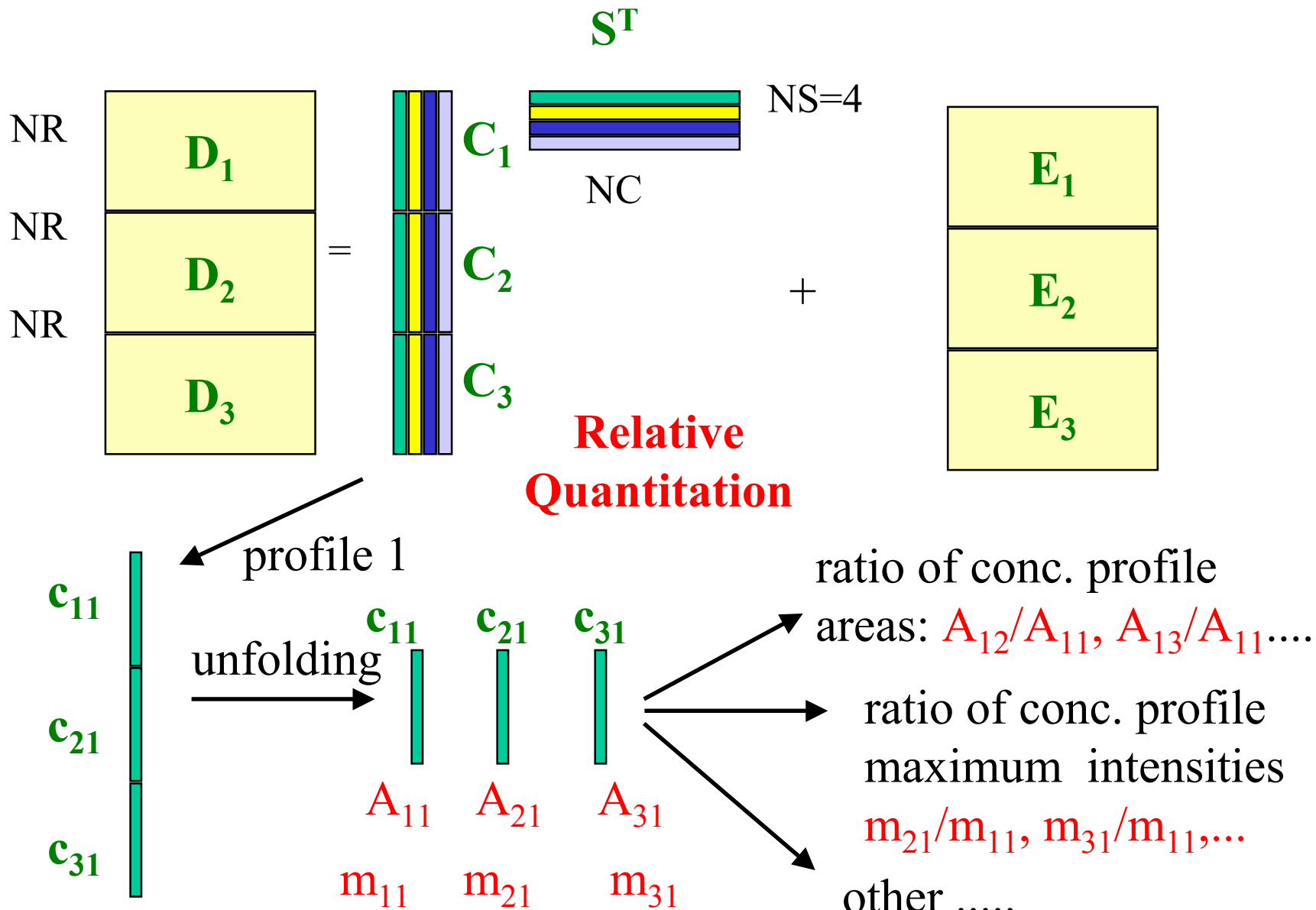
**appropriate experimental design of experiments included in augmented matrices may allow total resolution!**



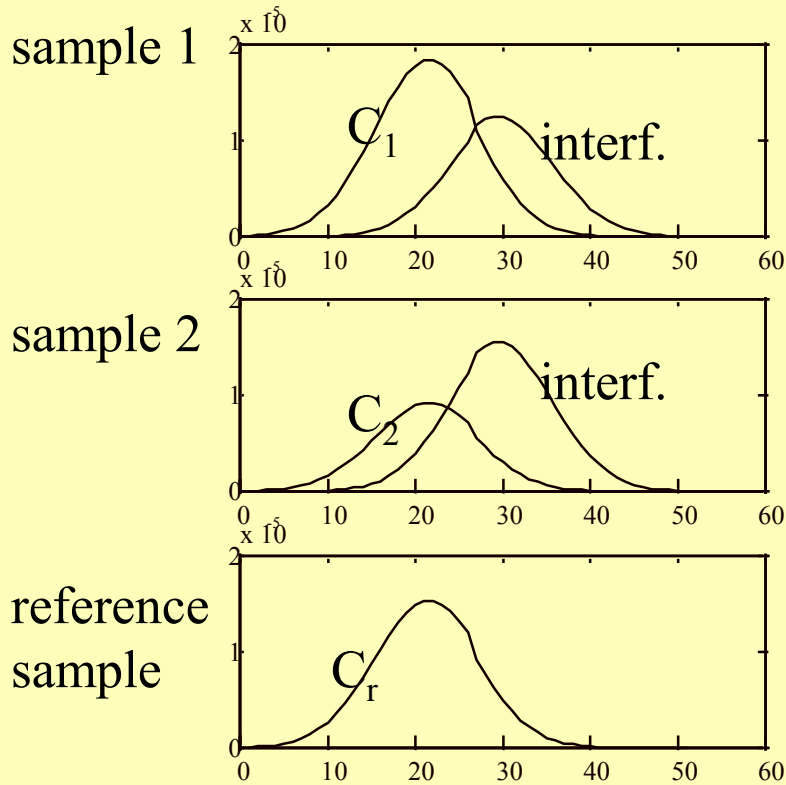
# Non-trilinear data: spectra recovery

Species	TLD	ALS (cos)	ALS (sin)
1	complex	0,9984	0,0567
2	complex	0,9997	0,0246
3	complex	1	0,008
4	complex	1	0,008

# Quantitative MCR-ALS for three-way data



# Recovery of quantitative information



- **Relative Quantitation**

reference  $C_r$  unknown

$$\text{Rel Conc}(C_1) = \text{Area}(C_1) / (\text{Area}(C_r))$$

$$\text{Rel Conc}(C_2) = \text{Area}(C_2) / (\text{Area}(C_r))$$

- **Absolute Quantitation**

reference  $C_r$  known

$$\text{Conc}(C_1) = [\text{Area}(C_1) / \text{Area}(C_r)] \text{ Conc}(C_r)$$

$$\text{Conc}(C_2) = [\text{Area}(C_2) / \text{Area}(C_r)] \text{ Conc}(C_r)$$

# Trilinear data: quantitative recovery

Species	Matrix	theoretical	TLD	ALS
1	2	0,5	0,5	0,5
	3	1,2	1,2	1,2
	4	0,7	0,7	0,7
2	2	0,8	0,85	0,84
	3	0,5	0,48	0,5
	4	0,66	0,67	0,67
3	2	1,87	1,85	1,87
	3	1,25	1,24	1,25
	4	0,62	0,62	0,62
4	2	0,8	0,82	0,81
	3	1,2	1,21	1,2
	4	0,5	0,5	0,5

# Non-trilinear data: quantitative recovery

Species	Matrix	theoretical	ALS
1	2	0,61	0,55
	3	0,81	0,84
	4	0,38	0,39
2	2	1,34	1,39
	3	0,34	0,31
	4	0,18	0,17
3	2	2,13	2,2
	3	1	1,07
	4	0,27	0,25
4	2	0,68	0,68
	3	0,27	0,26
	4	0,4	0,41

# Determination of triphenyltin in sea-water by excitation-emission matrix fluorescence and multivariate curve resolution

A method for the determination of triphenyltin (TPhT) in sea-water is proposed:

- 1) Solid phase extraction (SPE) of sea-water samples;
- 2) Reaction with a fluorogenic reagent (flavonol in a micellar medium);
- 3) Excitation-emission fluorescence measurements (giving an EEM data matrix);
- 4) MCR-ALS analysis of EEM data matrices
- 5) Quantitation of TPhT

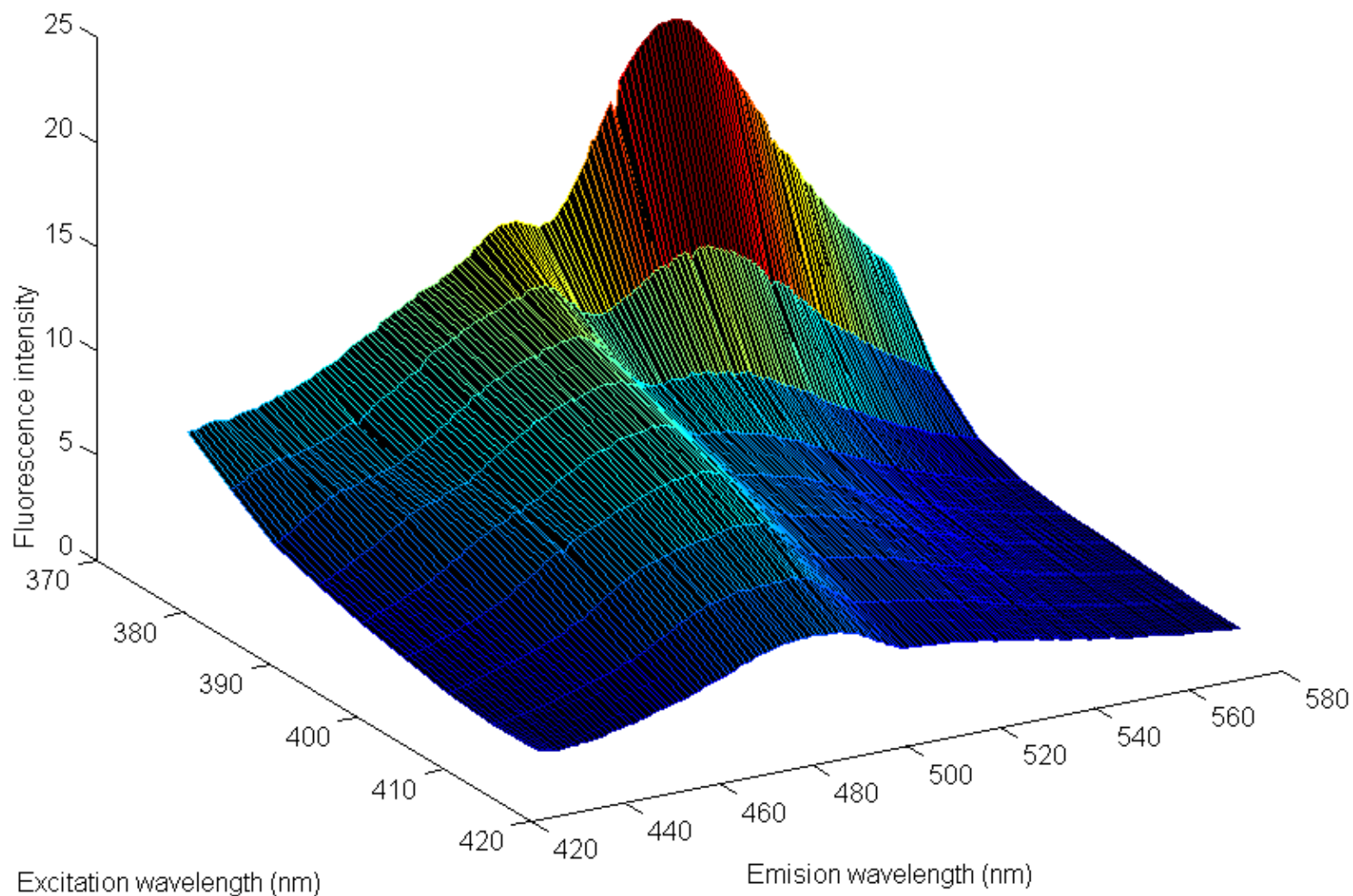
*J.Saurina, C.Leal, R.Compañó, M.Granados, R.Tauler and M.D.Prat.  
Analytica Chimica Acta, 2000, 409, 237-245*

# Determination of triphenyl in sea-water by excitation-emission matrix fluorescence and multivariate curve resolution.

Difficulties were:

- low concentrations of TPht (ng/l)
- strong background (fulvic acids) emission
- strong reagent emission
- lack of selective emission/excitation wavelengths
- to have sea-water TPhT standards available

# Excitation-Emission spectra for an unknown sea-water sample





# MCR-ALS resolution of EEM data

$$\begin{array}{c}
 \text{EEM} \\
 \mathbf{D}_{\text{aug}} \\
 \begin{array}{c}
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},n}
 \end{array}
 \begin{array}{c}
 \mathbf{U} \\
 \mathbf{S} \\
 \mathbf{R} \\
 \mathbf{B}
 \end{array}
 \end{array}
 =
 \begin{array}{c}
 \text{emission} \\
 \mathbf{Y}_{\text{aug}} \\
 \begin{array}{c}
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},n}
 \end{array}
 \begin{array}{c}
 \mathbf{Y}_{\text{U}} \\
 \mathbf{Y}_{\text{S}} \\
 \mathbf{Y}_{\text{R}} \\
 \mathbf{Y}_{\text{B}}
 \end{array}
 \end{array}
 \begin{array}{c}
 \text{excitation} \\
 \mathbf{X}^{\text{T}} \\
 \boxed{\mathbf{X}^{\text{T}}} \\
 \lambda_{\text{ex},1} \quad \lambda_{\text{ex},m} \\
 + \\
 \lambda_{\text{ex},1} \quad \lambda_{\text{ex},m}
 \end{array}
 +
 \begin{array}{c}
 \text{noise} \\
 \mathbf{E}_{\text{aug}} \\
 \begin{array}{c}
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},m} \\
 \lambda_{\text{em},1} \\
 \lambda_{\text{em},n}
 \end{array}
 \begin{array}{c}
 \mathbf{E}_{\text{U}} \\
 \mathbf{E}_{\text{S}} \\
 \mathbf{E}_{\text{R}} \\
 \mathbf{E}_{\text{B}}
 \end{array}
 \end{array}
 \begin{array}{c}
 \lambda_{\text{ex},1} \quad \lambda_{\text{ex},m}
 \end{array}
 \end{array}$$

U unknown sea water; S TPhT pure standard;  
 R reagent (flavonol); B sea-water background (fulvic acids)

# MCR-ALS resolution of EEM data

## *a) Model:*

$$[\mathbf{U};\mathbf{S};\mathbf{R};\mathbf{B}] = \mathbf{D}_{\text{aug}} = \mathbf{Y}_{\text{aug}}\mathbf{X}^T + \mathbf{E}_{\text{aug}}$$

## *b) Resolution:*

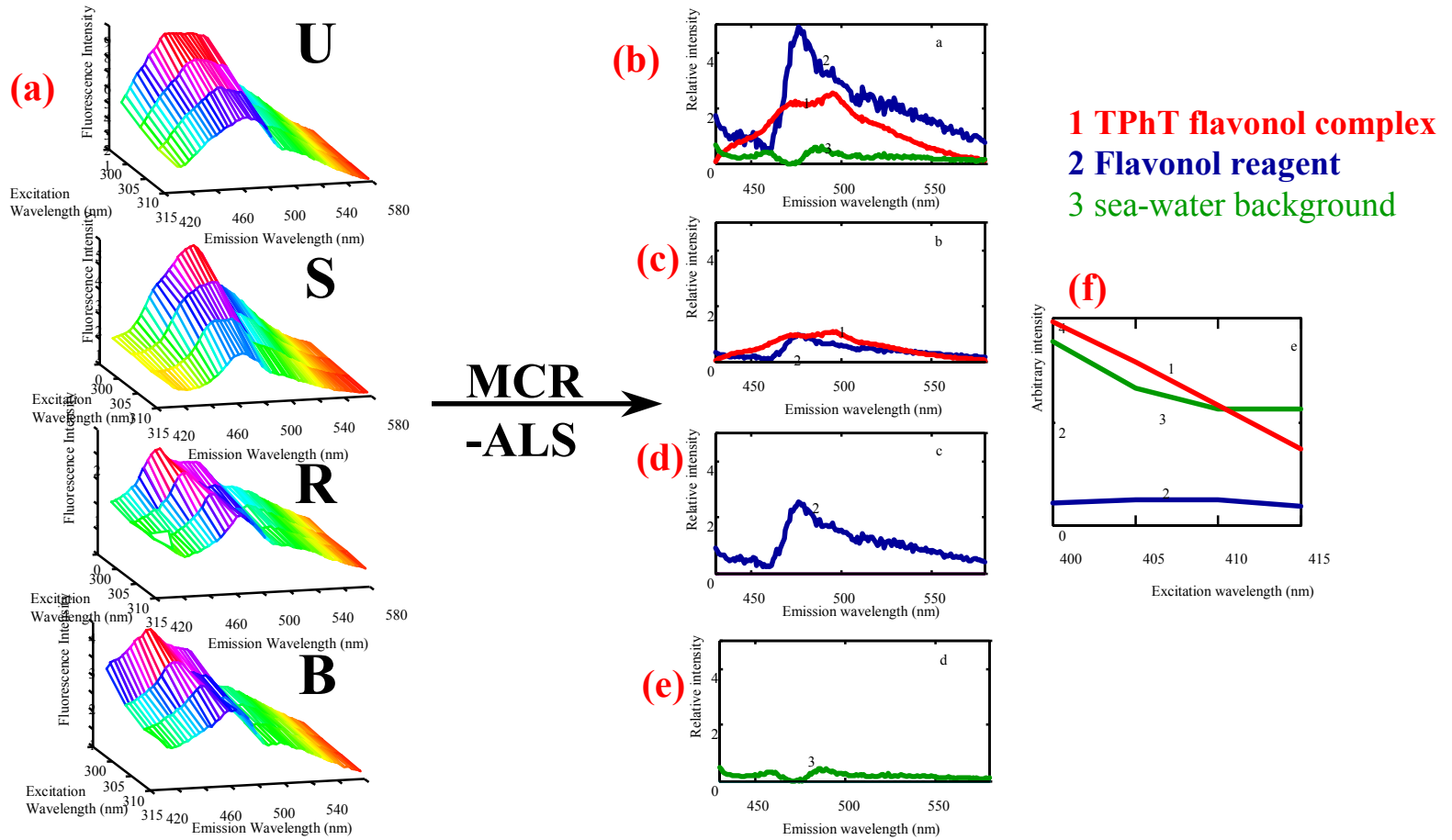
(emission)  $\mathbf{Y}_{\text{aug}} = \mathbf{D}_{\text{aug}} (\mathbf{X}^T)^+$  Constraints:  
- non-negativity

(excitation)  $\mathbf{X}^T = (\mathbf{Y}_{\text{aug}})^+ \mathbf{D}_{\text{aug}}$  - trilinearity

## *c) Quantitation:*

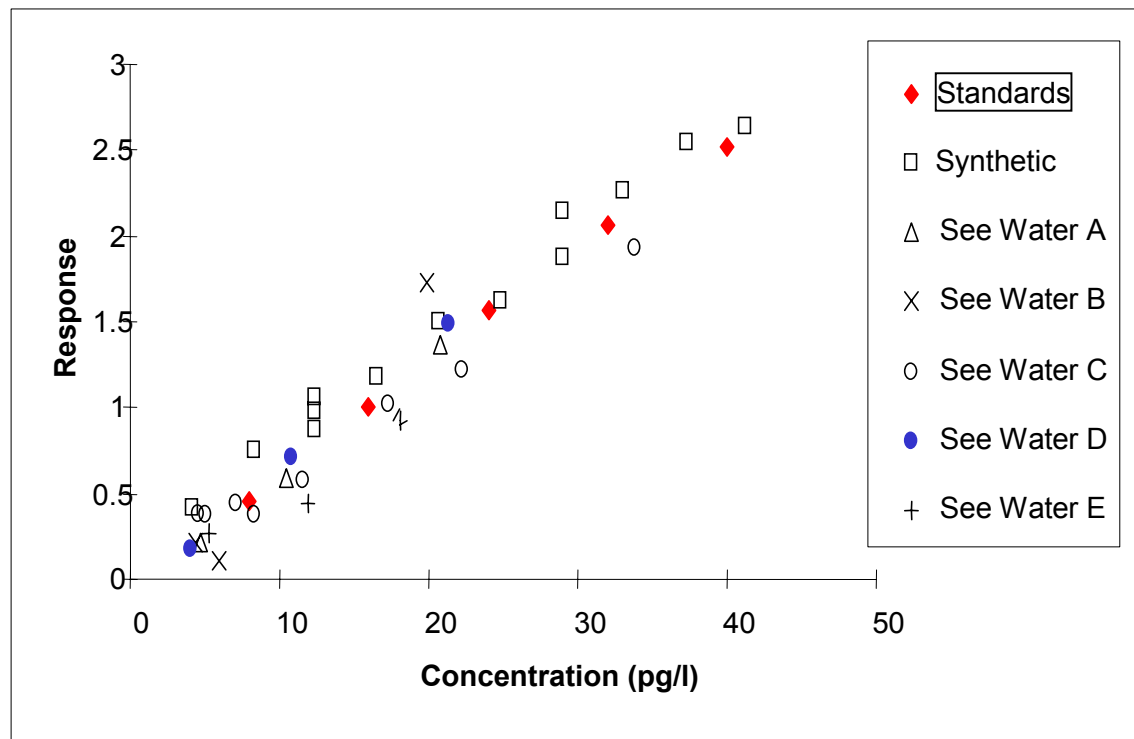
$$c_{TPhT,U} = [Area(y_{TPhT,U}) / Area(y_{TPhT,S})] c_{TPhT,S}$$

# MCR-ALS resolution of [U;S;R;B] augmented matrix



- a)** 3-D plots of the EEM fluorescence of the unknown sample U, standard S, flavonol reagent R and sea-water background B;
- b)** emission spectra for the unknown sea-water sample; **c)** emission species spectra for the standard;
- d)** emission species spectra for flavonol reagent; **e)** emission species spectra for sea-watere background;
- f)** excitation spectra

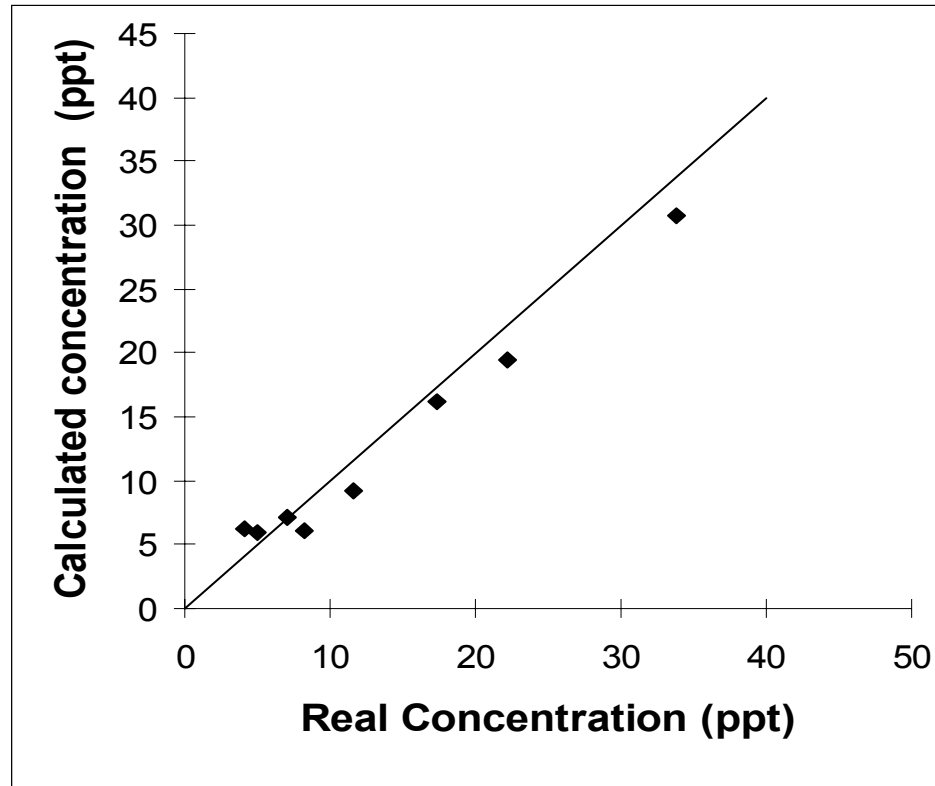
# MCR-ALS resolution/quantitation of EEM data



Plot of the emission profiles areas for TPhT species in standards, synthetic and sea-water samples respect the analyte concentration

# Comparison between 'true' and MCR-ALS calculated TPhT concentrations in sea-water samples

**Quantitation:**  $c_U = [\text{Area}(y_U) / \text{Area}(y_S)] c_S$



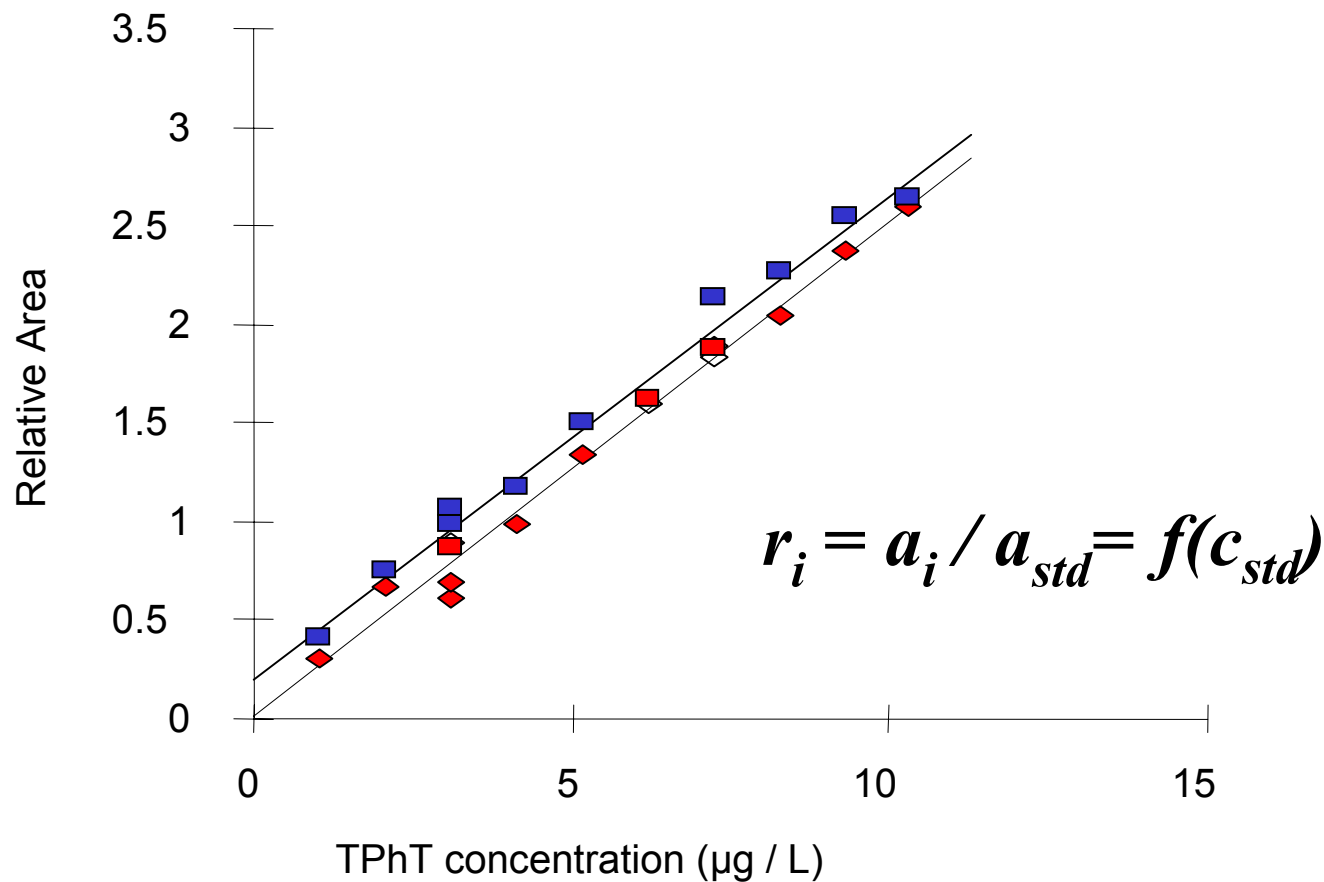
*overall prediction errors were always below 13%!*

## FIGURES OF MERIT IN SECOND ORDER MULTIVARIATE CURVE RESOLUTION

- From MCR-ALS resolution of the pure response profiles of the analyte in different known and unknown mixtures (data matrices), a *Calibration Curve* is built.
- Figures of merit such as *Limit of Detection, Sensitivity, Precision and Accuracy* are calculated from the calibration curve

*like in univariate calibration!*

# Building the Calibration Curve and Sensitivity



◆ Approach (a) [U;S2;R]  $r_i = 0.260 c_i + 0.014$  ( $r = 0.998$ )

■ Approach (b) [U1;U2;U3;U4;U5;U6;U7;U8;U9;U10;U11;U12;S2;R;B]  
 $r_i = 0.244 c_i + 0.201$  ( $r = 0.987$ )

## Precision:

$$s_R = \sqrt{\frac{\sum_{i=1}^n (\hat{r}_i - \bar{r})^2}{n-1}}$$

(a) and (b)  $s_R = 0.0404$

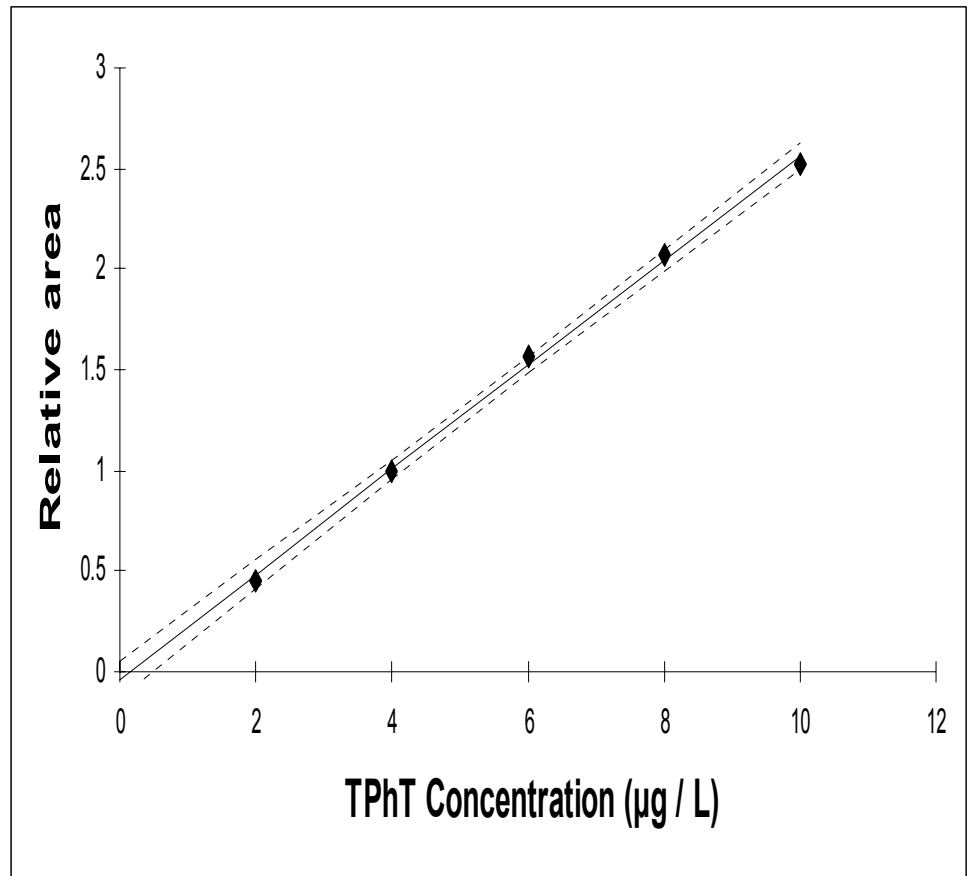
## Limit of detection

$$\text{LOD} = + t s_R / b \left( 1/m + 1/n + \frac{((r_i - \bar{r}) / b)^2 / \Sigma(c_i - \bar{C})^2}{1} \right)^{1/2}$$

(a) and (b)  $\text{LOD} = 0.7 \mu\text{g l}^{-1}$

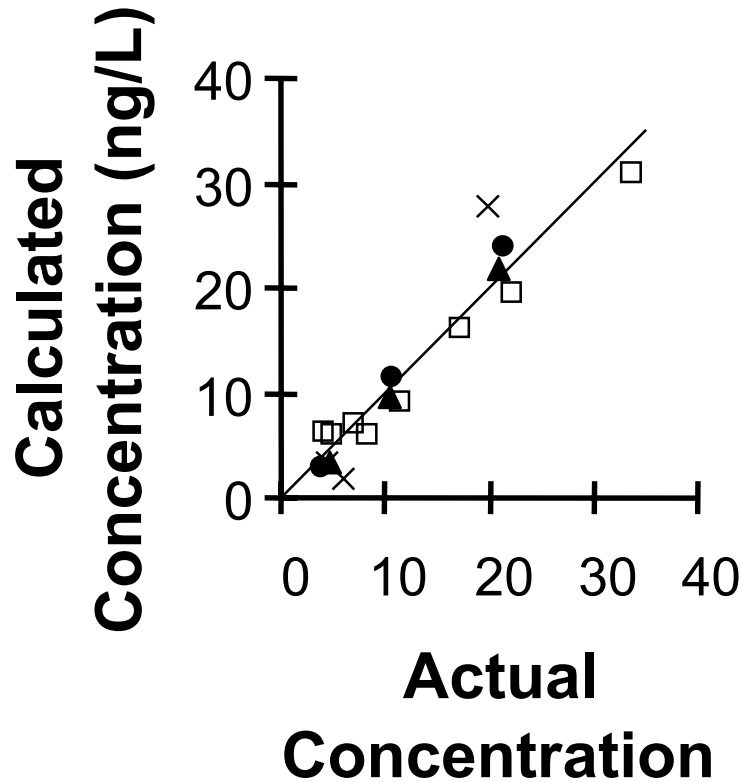
## Precision bands

$$\pm s_R t \left( 1/m + 1/n + (r_i - \bar{r})^2 / \Sigma(c_i - \bar{C})^2 \right)^{1/2}$$





# *Accuracy of the method in the prediction of TPhT in real samples*



- ▲ Sea Water A
- × Sea Water B
- Sea Water C
- Sea Water D

*overall prediction error*

$$\text{Error}(\%) = \frac{\sqrt{\sum_{i=1}^{\text{Samples}} (c_i - \hat{c}_i)^2}}{\sqrt{\sum_{i=1}^{\text{Samples}} (c_i)^2}} \times 100$$

***Error % = 5.5 % for strategy (A)***  
***Error % = 12.7 % for strategy (B)***

# Solving matrix effects

Three strategies were compared for the recovery of the analyte response in the sea-water samples:

- i. using pure standards*
- ii. using sea-water standards*
- iii. using the standard addition method*

*J.Saurina and R.Tauler, The Analyst, 2000, in press*

# Standard addition strategy:

For each unknown sample, MCR-ALS is applied to the following augmented matrices (i.e A4, the same for the other A1, A2, A3, A5 and A6)

## *augmented matrices*

## *identification*

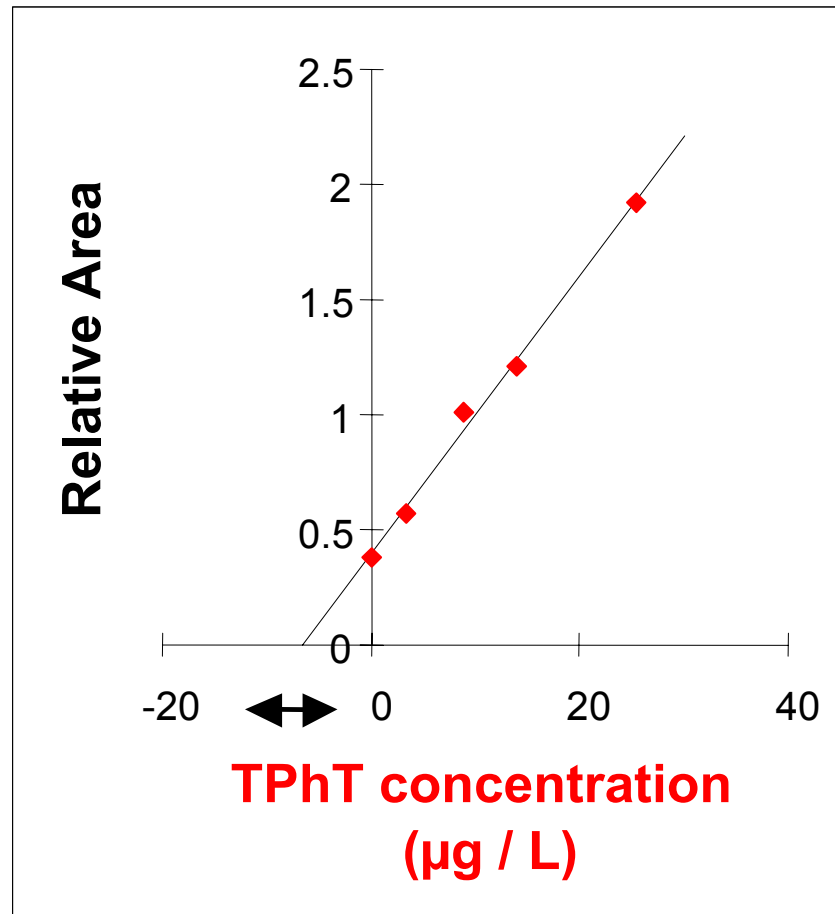
[A4;S2;R;B]	=>	A4 unknown sample
[A4SA1;S2;R;B]	=>	A4SA1 = A4 + 0.20 $\mu\text{g l}^{-1}$ TPhT
[A4SA2;S2;R;B]	=>	A4SA2 = A4 + 0.75 $\mu\text{g l}^{-1}$ TPhT
[A4SA3;S2;R;B]	=>	A4SA3 = A4 + 1.05 $\mu\text{g l}^{-1}$ TPhT
[A4SA4;S2;R;B]	=>	A4SA4 = A4 + 1.87 $\mu\text{g l}^{-1}$ TPhT
[A4SA5;S2;R;B]	=>	A4SA5 = A4 + 3.30 $\mu\text{g l}^{-1}$ TPhT
[A4SA6;S2;R;B]	=>	A4SA6 = A4 + 4.52 $\mu\text{g l}^{-1}$ TPhT
[A4SA7;S2;R;B]	=>	A4SA7 = A4 + 7.42 $\mu\text{g l}^{-1}$ TPhT

**S2** EMM response matrix of an standard of TPhT

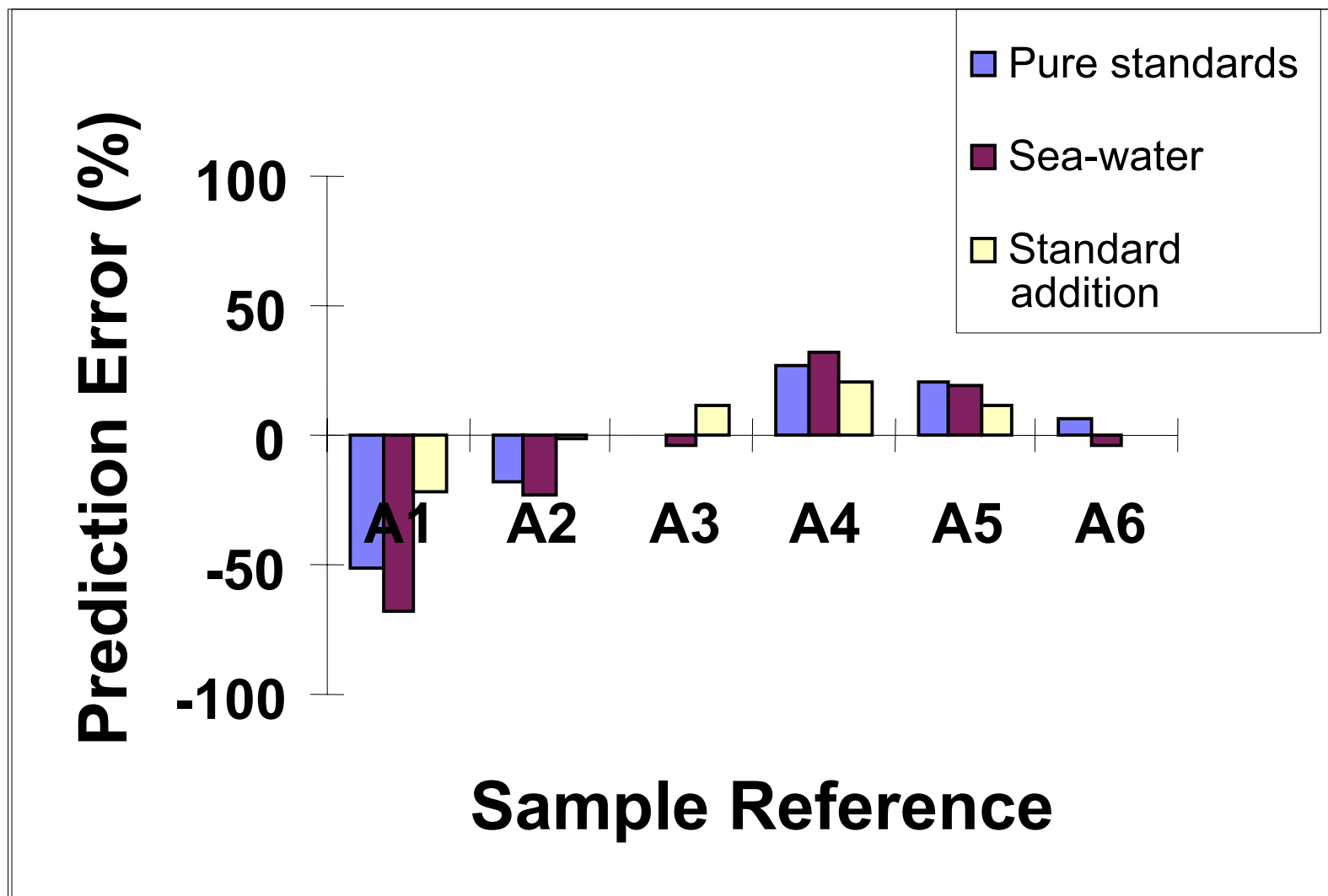
**R** EMM response matrix of the reagent

**B** EMM response matrix of the background

# Standard addition calibration graph in a sea-water analyte determination (sea-water sample A4)



# Prediction errors in the determination of TPhT in sea-water samples A1-A6 using MCR-ALS and three calibration approaches:



# Provisory conclusions on the use of MCR-ALS for quantitative three-way data analysis

- ✓ Quali- and Quantitative information may be recovered simultaneously using MCR-ALS for three-way data
- ✓ Calculation of figures of merit is possible from resolved profiles (using same methods as in univariate calibration)
- ✓ The combination of standard addition with multivariate curve resolution method improved the accuracy of predictions in the presence of matrix effects.
- ✓ Limitations will appear for analytes contributing very little to the whole signal and for systems with a high chemical rank

# Acknowledgements

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University of Barcelona**

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Montse Vives, Susana Navea, Tarik Azzouz  
Alba Puigdomenech (NIR data set)